

Usage Cases of GBS

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Cornell CBSU Workshop

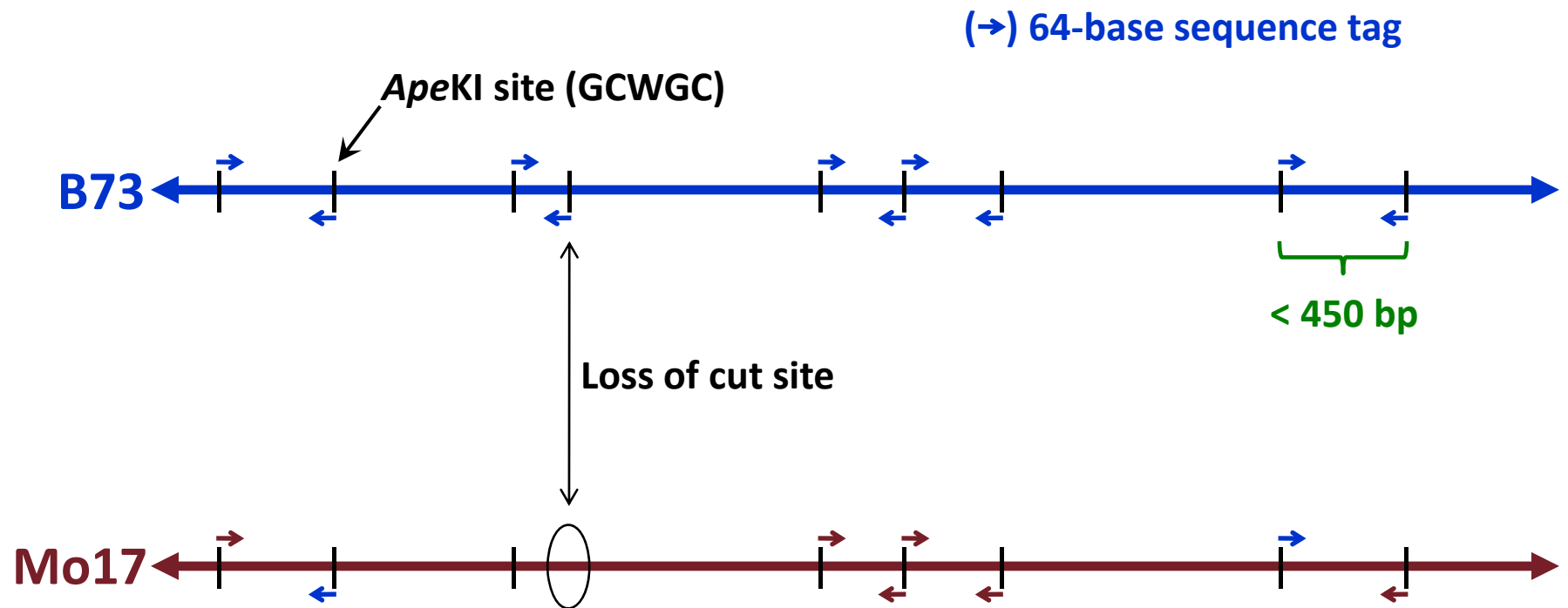
Oct 31-Nov 1, 2011

Some potential applications of GBS Data

- **Marker discovery**
- **Phylogeny/Kinship**
- **Linkage mapping of QTL in a biparental cross**
- **Fine-mapping QTL**
- **Bulked segregant analysis**
- **Genomic selection**
- **Genome Wide Association Studies (GWAS)**
- **NAM-GWAS**
- **Improving reference genome assembly**

Marker Discovery

- GBS markers can be converted to SNPs or PCR assays of indels
- Develop SNP assays from polymorphic tags at same location
- Develop PCR primers from adjacent tags & hope for large indels



Phylogeny/Kinship

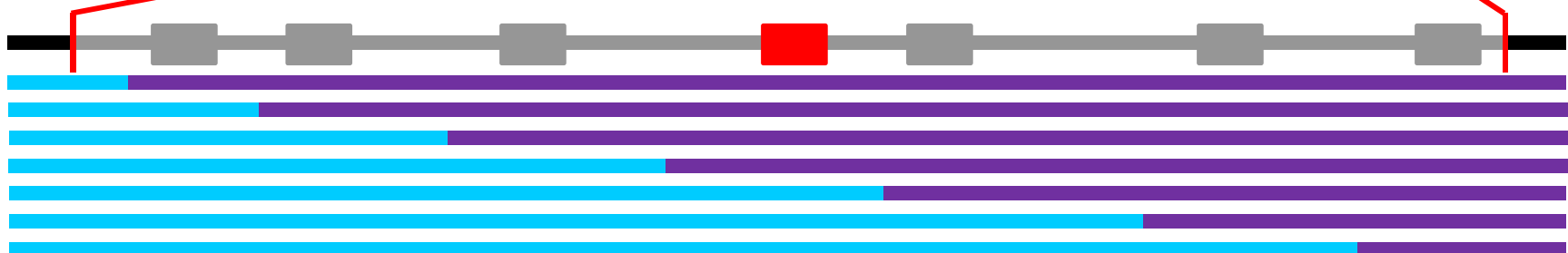
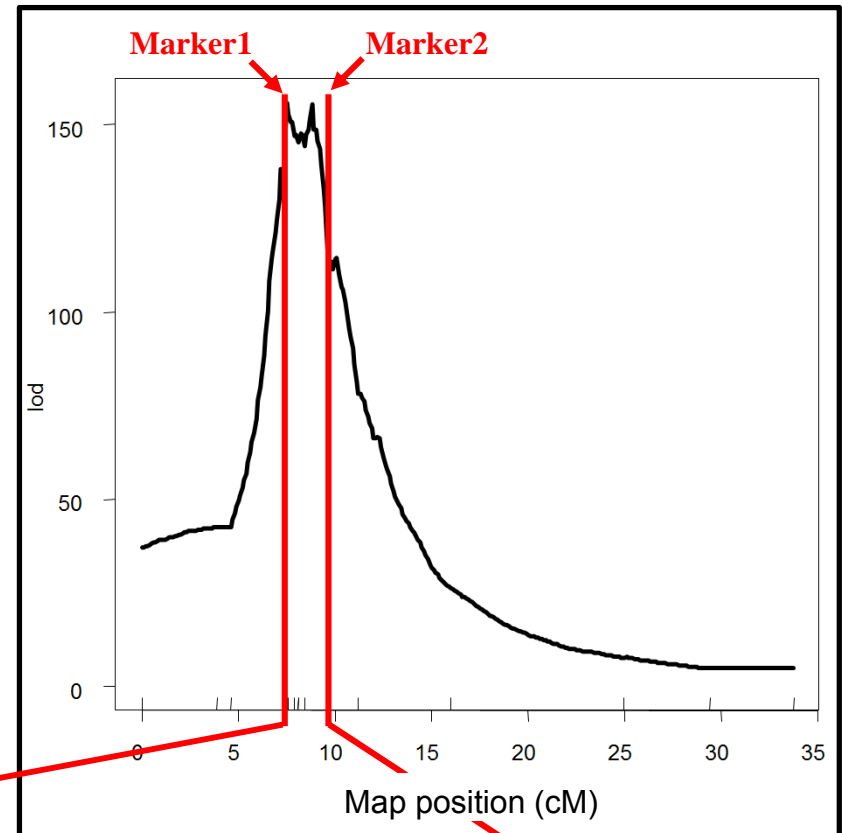
- **Missing data not an issue for estimating pairwise genetic distance or kinship**
 - Each pair of individuals has large, “random” sample of markers in common
- **Works really well even in non-model organisms**
 - Fei Lu’s previous talk on switchgrass
- **Principle Coordinates Analysis better than Principle Components Analysis**
 - Uses distance matrix rather than every genotype
 - Missing data not an issue for Prin. Coord. Analysis
- **SNPs can be strongly affected by ascertainment bias**
 - Panel used to discover the SNPs can severely distort estimates of population genetic parameters (e.g., kinship, diversity)
 - Industry SNPs on the Maize 55K SNP chip an extreme example

Linkage mapping of QTL in a biparental cross

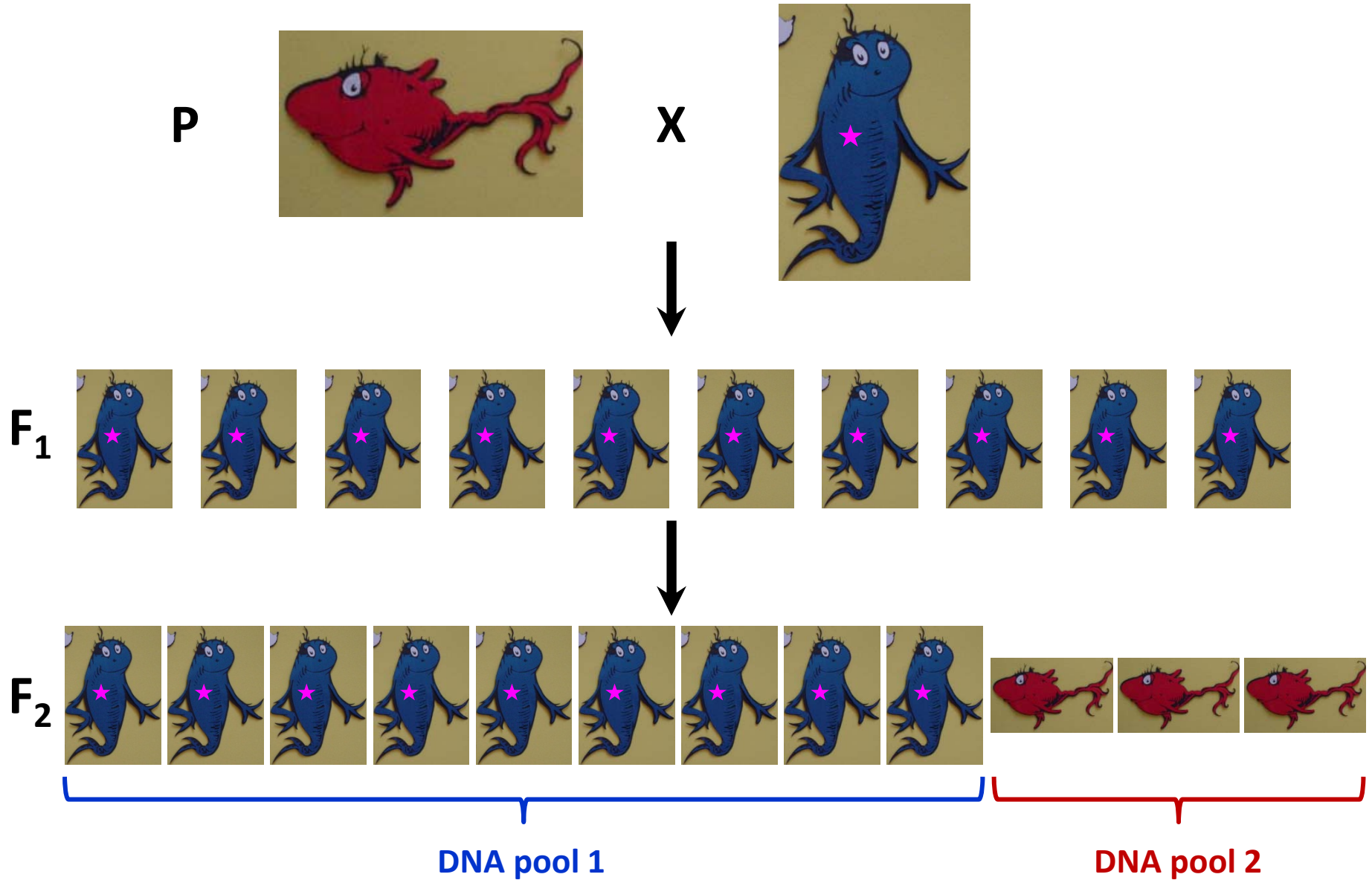
- In maize, we use the reference genome to order markers
- With *ApeKI*, too many markers for traditional software (MapMaker, JoinMap, R-QTL etc.)
- Filter for a smaller set of markers with high coverage
- Use *PstI* for fewer markers with higher coverage
- JoinMap can handle at least 3,000 markers
- Newer software?
 - MSTMap claims 10,000 – 100,000 markers
 - Others?

Fine mapping QTL

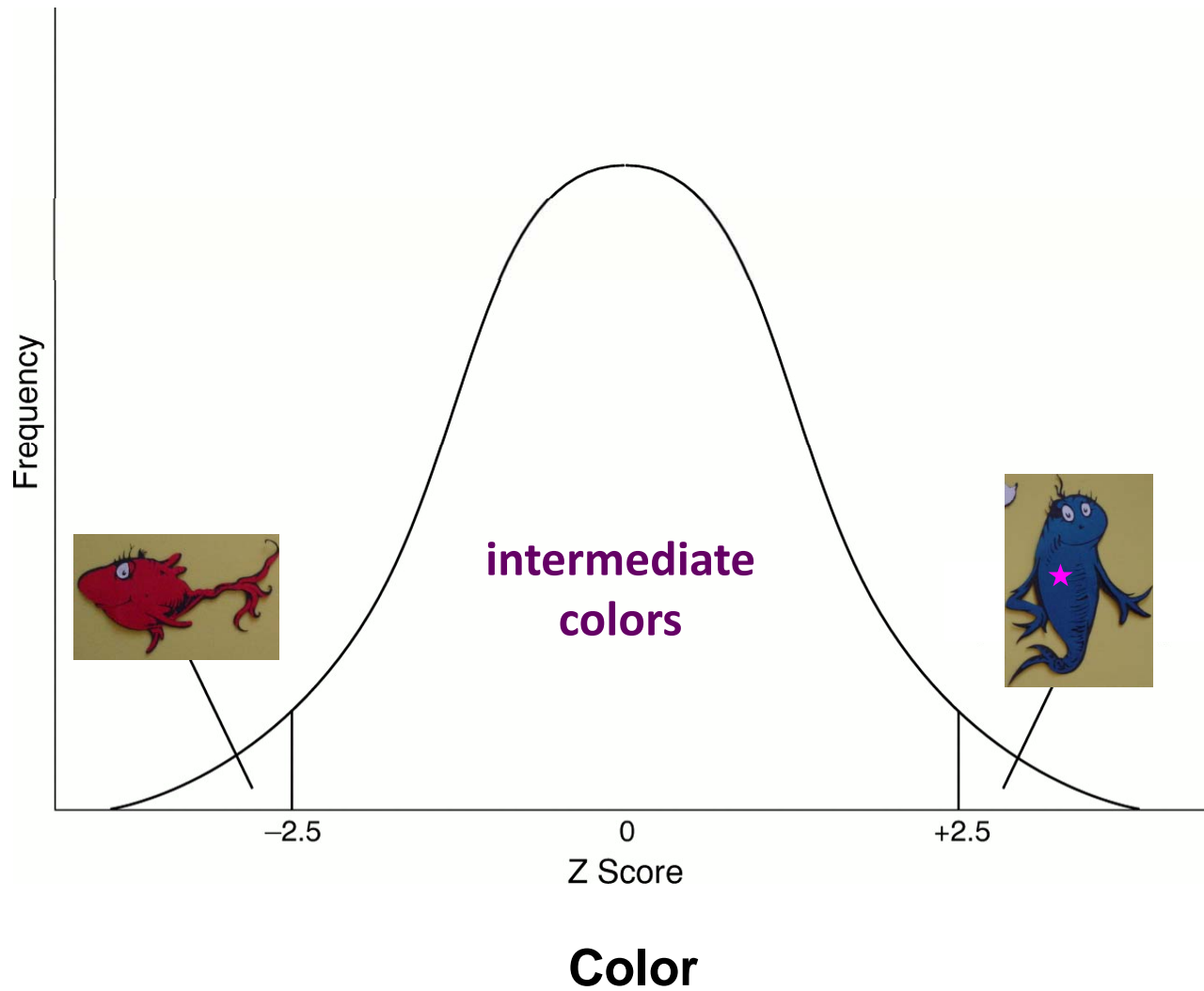
- Need to saturate interval containing QTL with markers
- GBS a good source of markers
- Also need to collect recombinants in the interval
- Near-isogenic lines (NILs) helpful (Mendelize)
- Good reference genome



Bulked Segregant Analysis



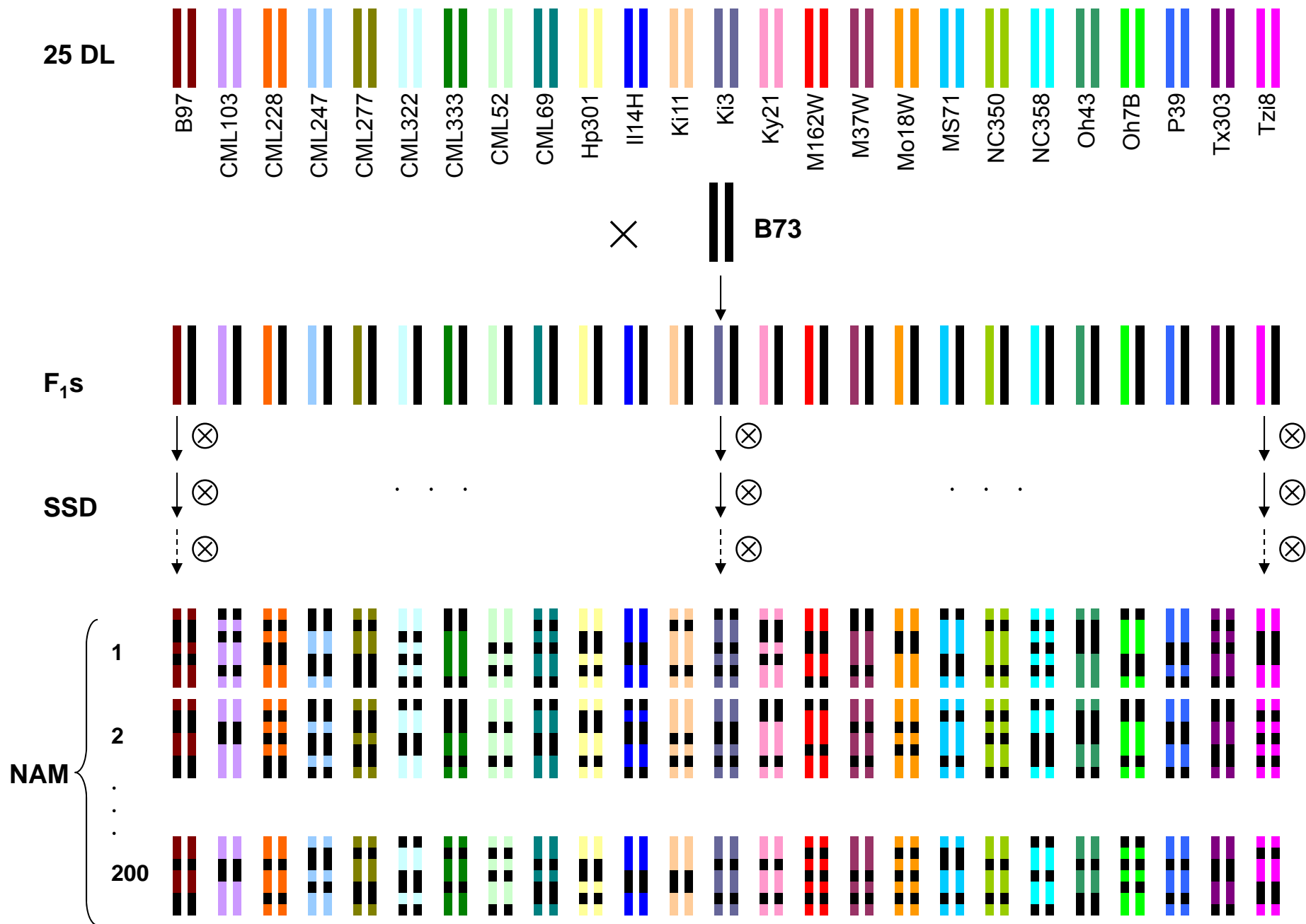
Bulked Segregant Analysis



Genomic Selection & GWAS

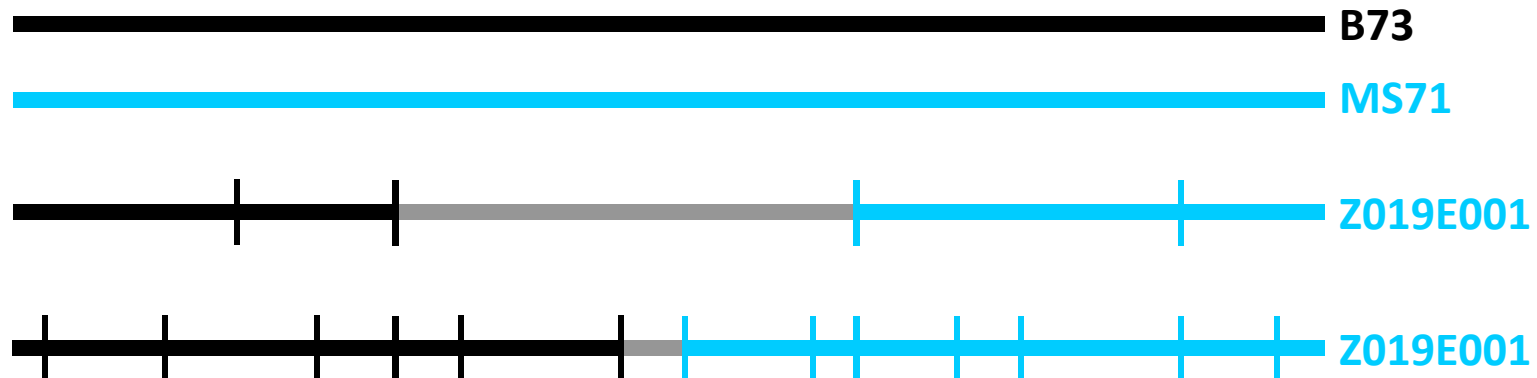
- **Complete data not required for genomic selection**
 - Closely linked markers in LD cover for each other
- **In contrast, missing data are more problematic for GWAS**
 - imputation necessary, but might cause spurious results
 - avoid false imputation of biologically missing regions
 - area of active research
- **In NAM-GWAS, imputation is much less of an issue**
 - NAM = “Nested Association Mapping” population

The maize NAM population was built for NAM-GWAS

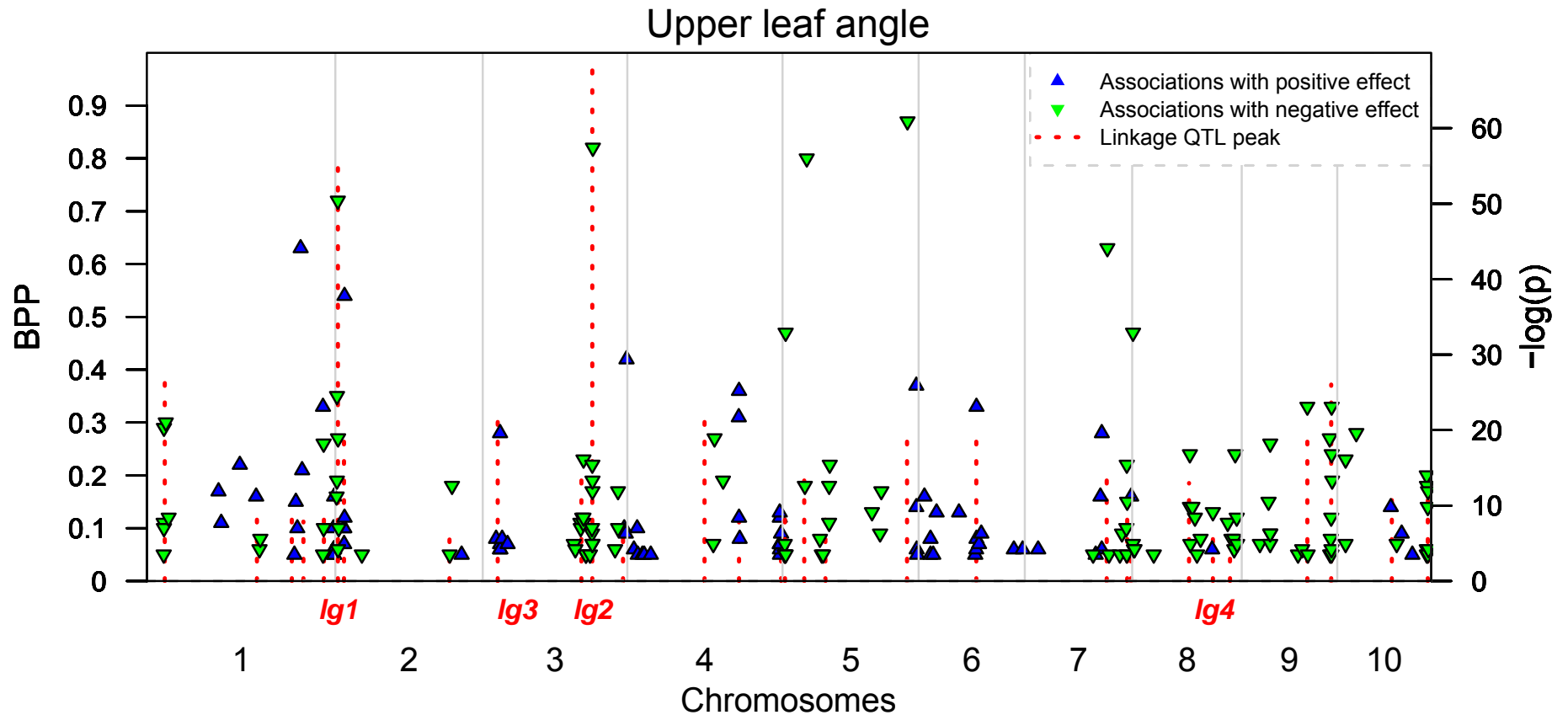


We are using GBS to pinpoint the location of cross overs in the NAM RILs

- B73 is the reference genome: complete knowledge
- Remaining NAM parents whole genome sequenced via Illumina at 4x coverage (paired end random sheared)
 - 26 million high quality SNPs
- Precise knowledge of crossover locations in NAM RILs allows us to more accurately project sequences of parents onto RILs:

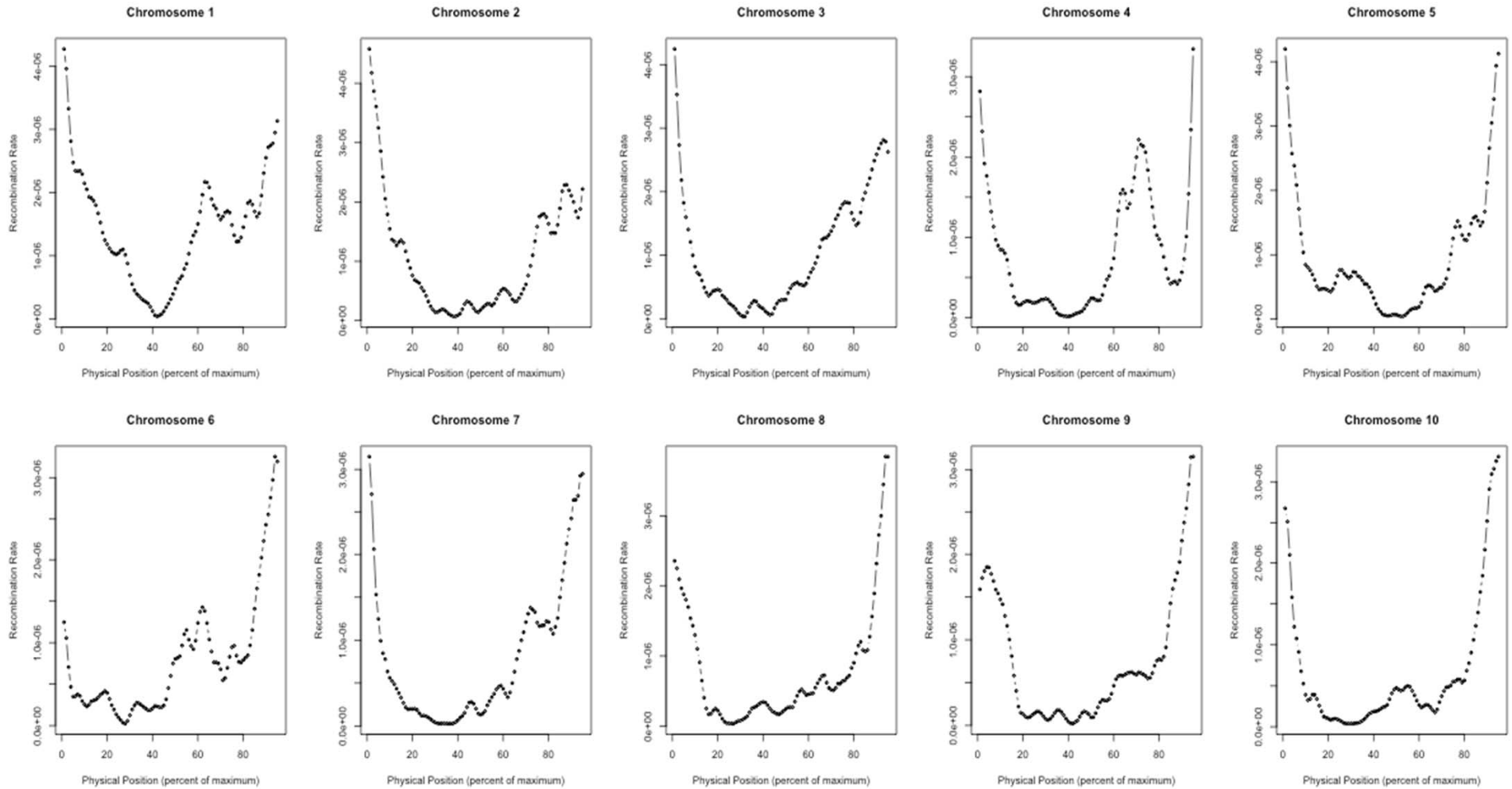


liguleless1 and *liguleless2* explain the two “biggest” leaf angle QTL



Tian, Bradbury, et al 2011 Nature Genetics

Recombination Rates for NAM from GBS Data



Peter Bradbury – USDA Scientist, Buckler lab, Cornell (unpublished)

The maize B73 reference genome: room for improvement?

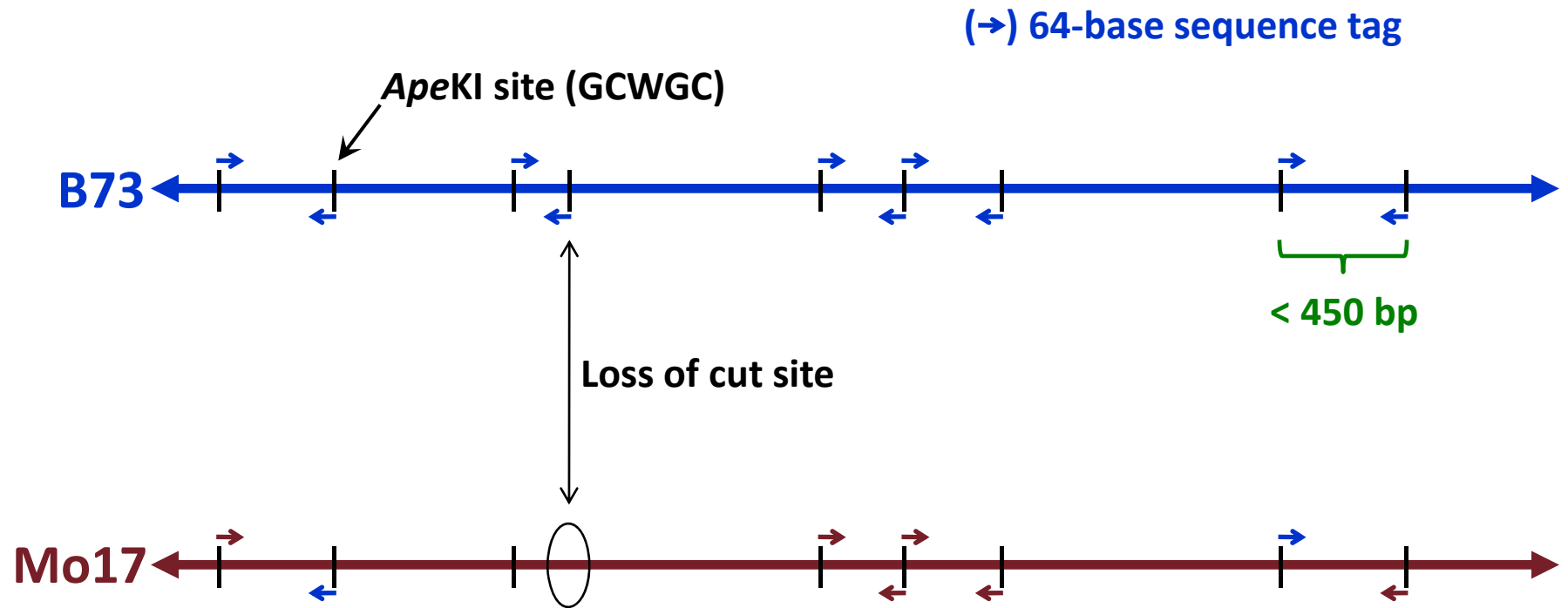
- 1) The B73 reference genome accurate for B73 but less so for other maize lines (*e.g.*, Mo17)
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- 3) Some large (multiple BAC) contigs could not be anchored
 - assigned to “chromosome 0”
 - 30 chr0 contigs in B73 RefGenV1
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- 4) Some regions of the genome are missing
 - ≈5% of B73 sequence is not in the B73 reference genome

The maize B73 reference genome: room for improvement?

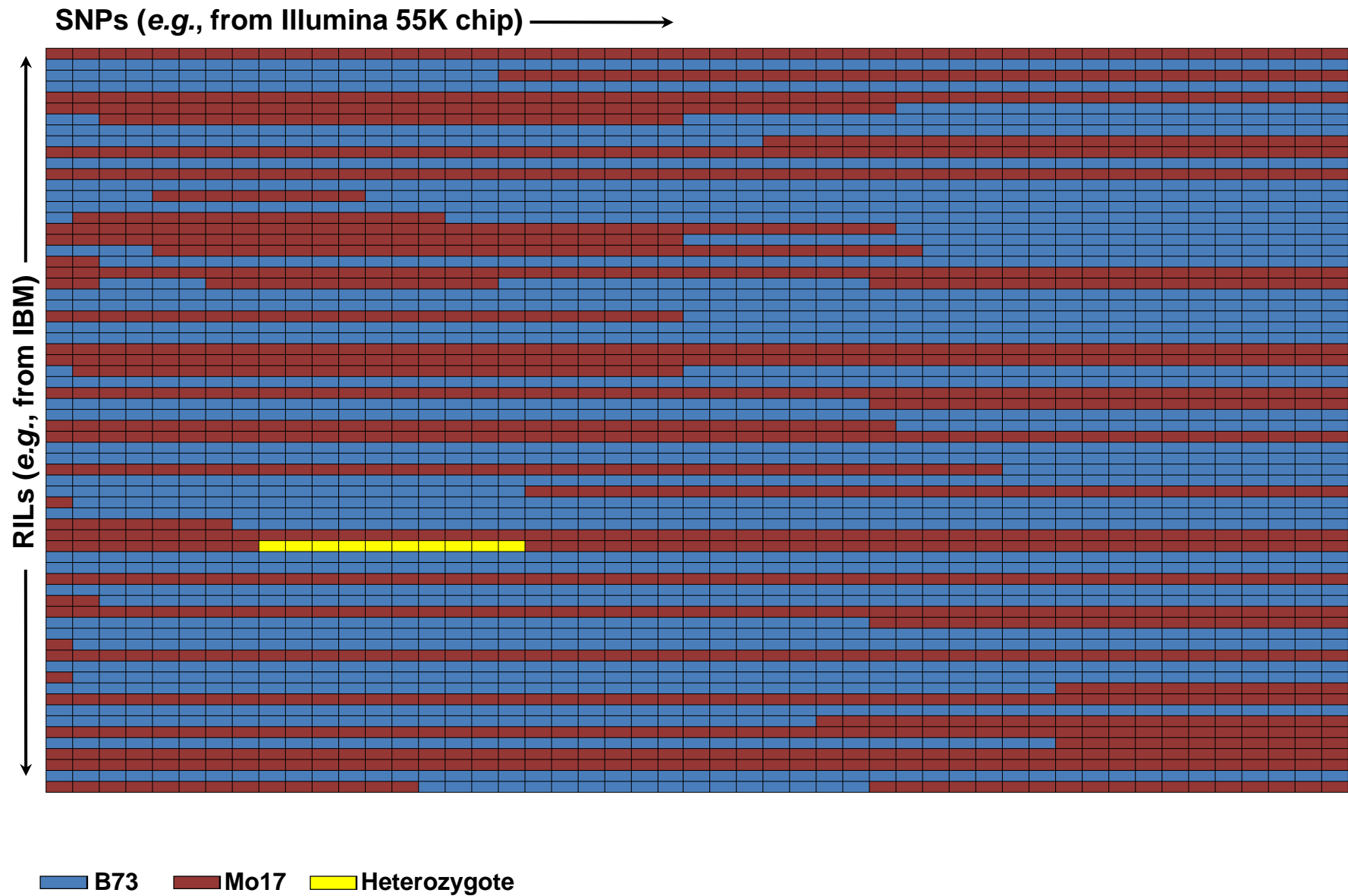
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Most tags can be mapped as individual alleles

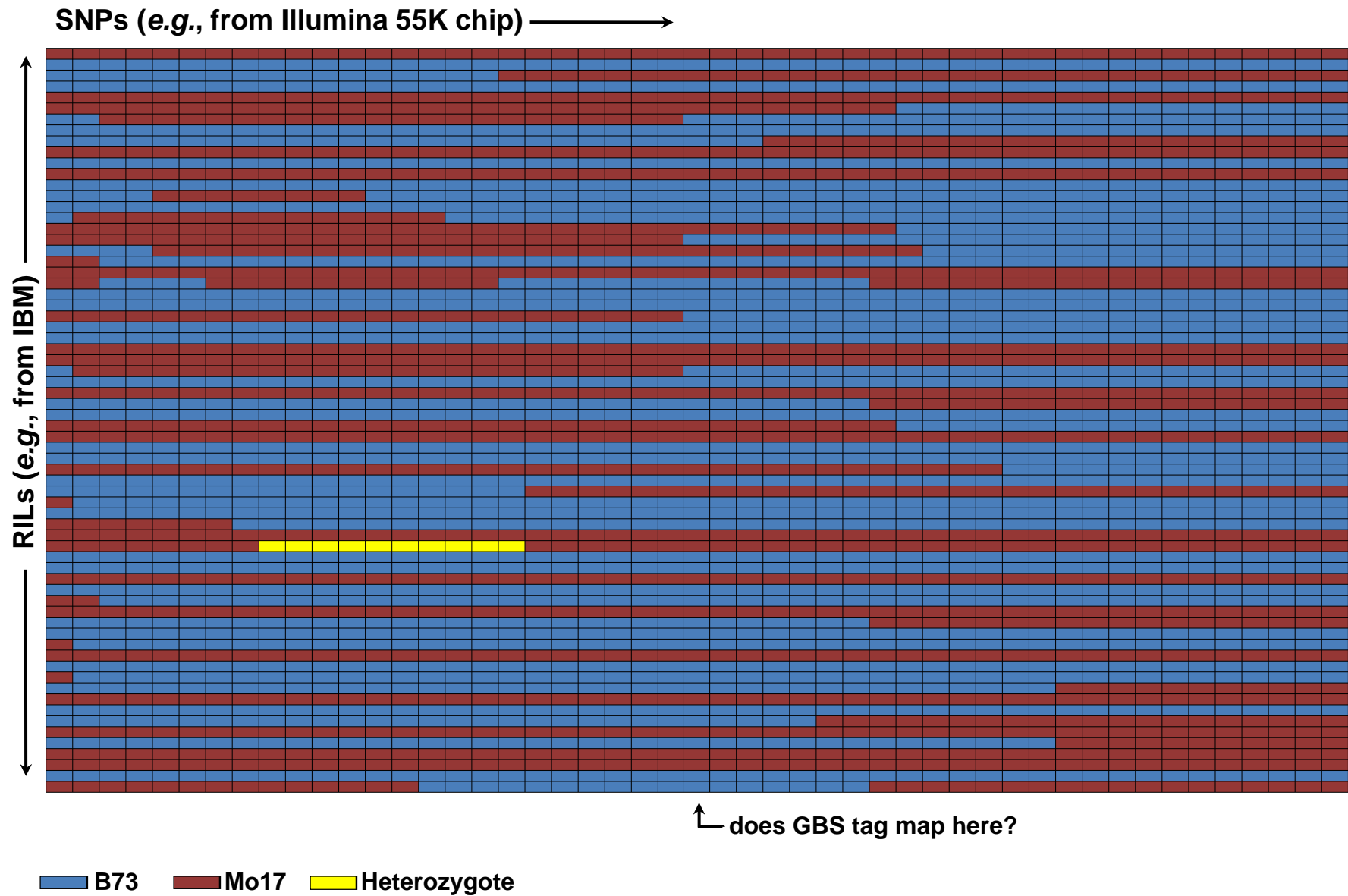
- In a biparental cross such as maize IBM (B73 x Mo17)
- Provided that they are polymorphic between the parents



Genetically mapping individual GBS alleles

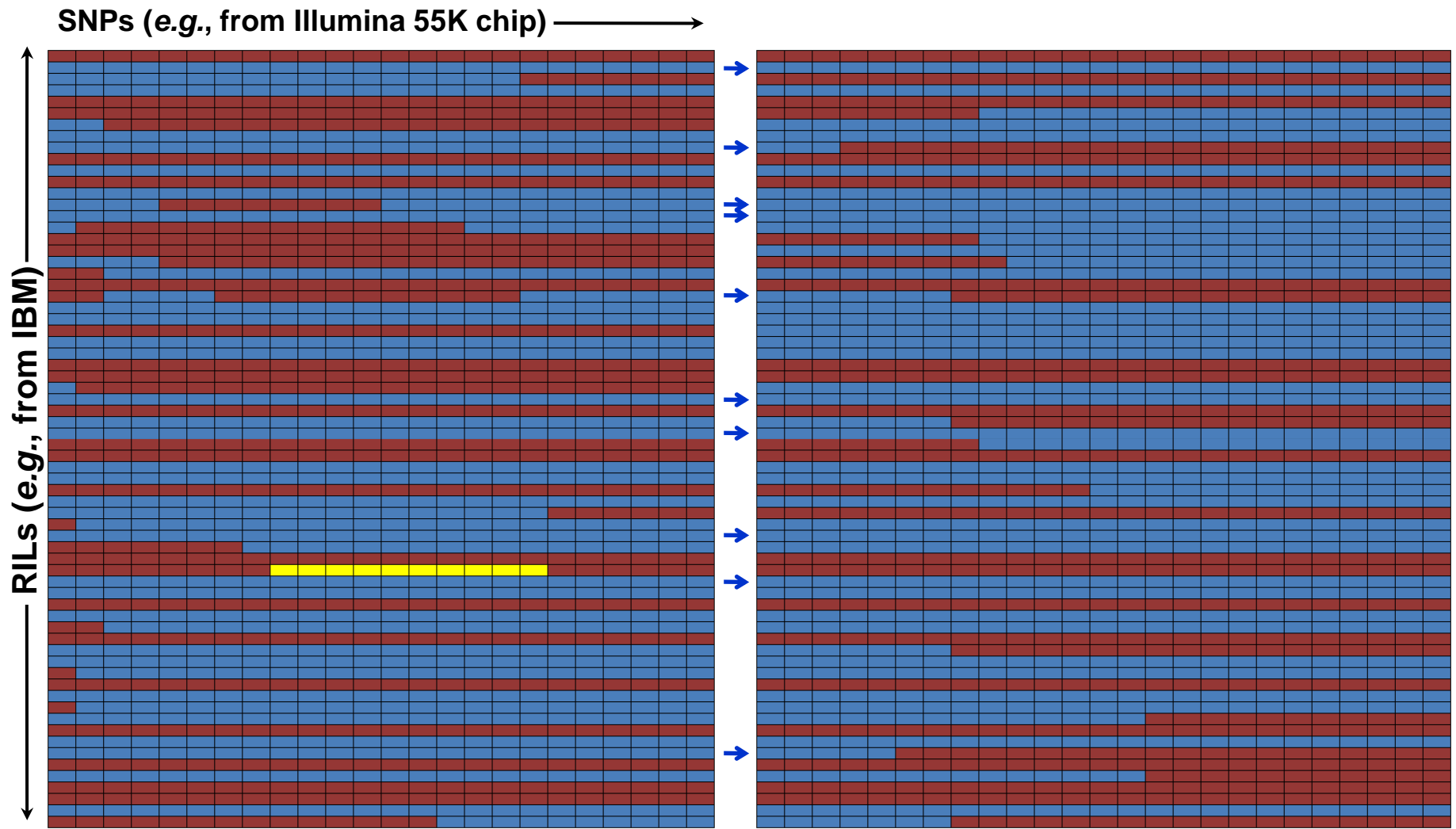


Genetically mapping individual GBS alleles



Genetically mapping individual GBS alleles

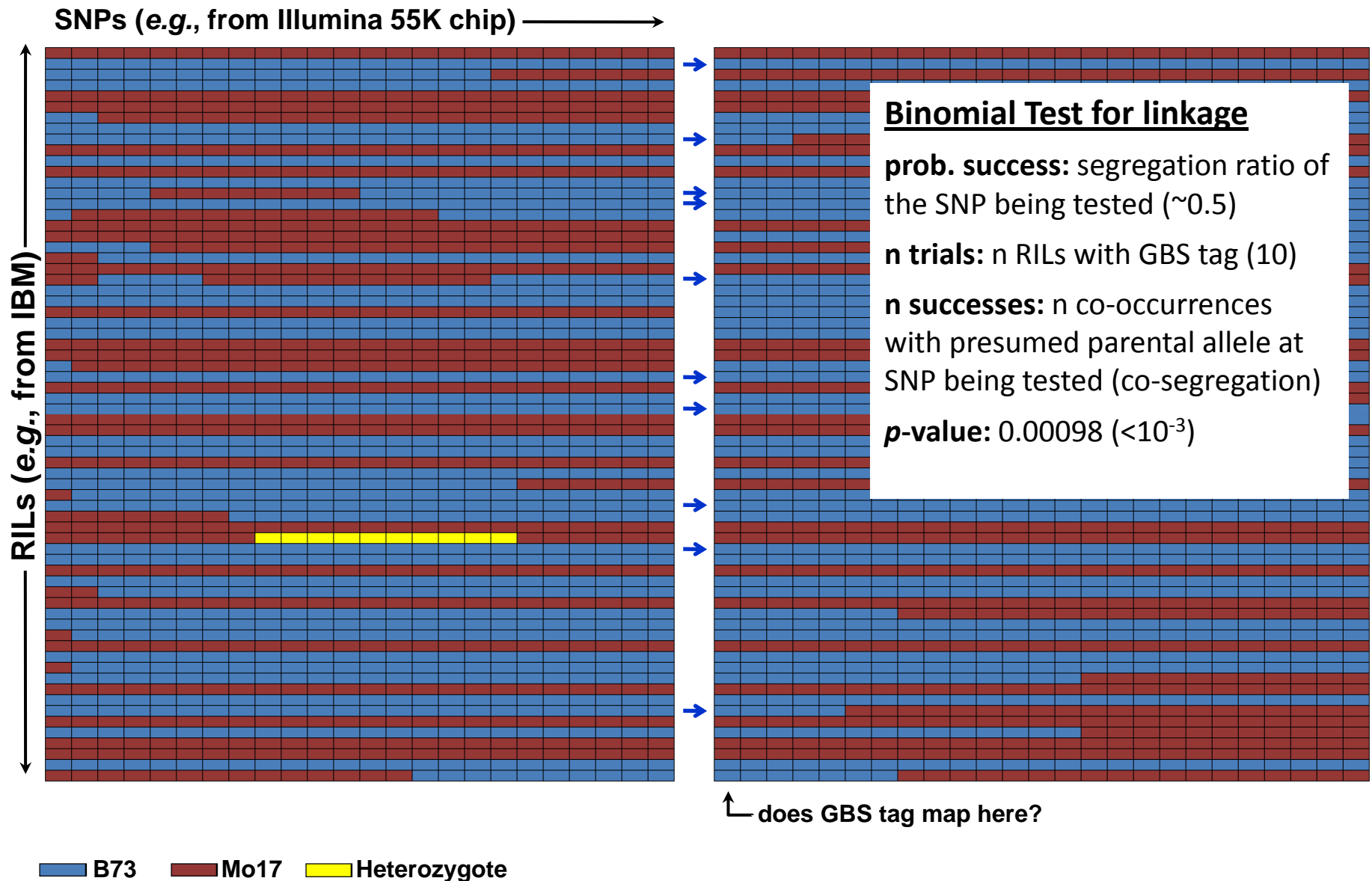
(→) 64-base sequence tag (GBS coverage ~0.4x)



■ B73 ■ Mo17 ■ Heterozygote

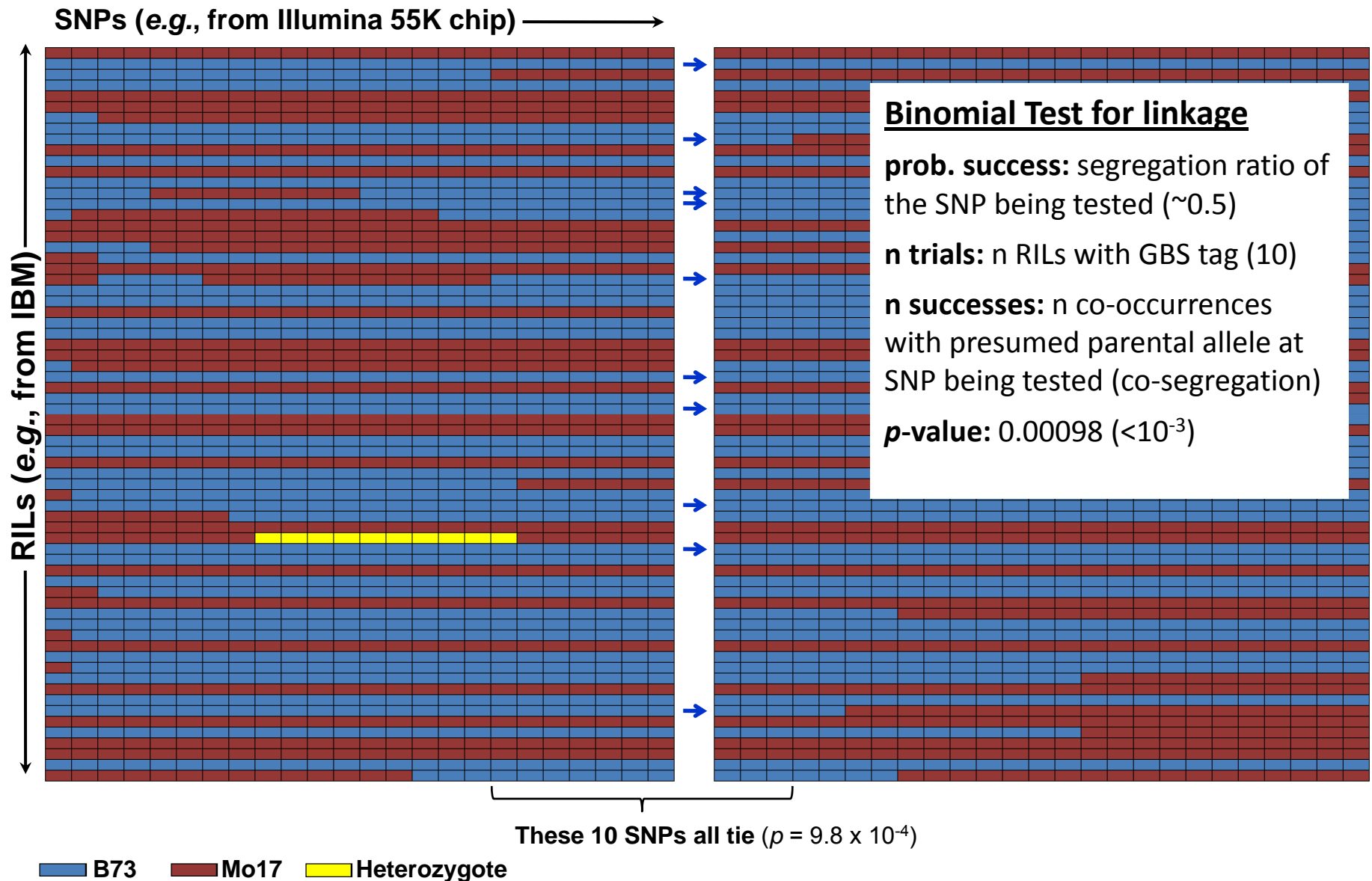
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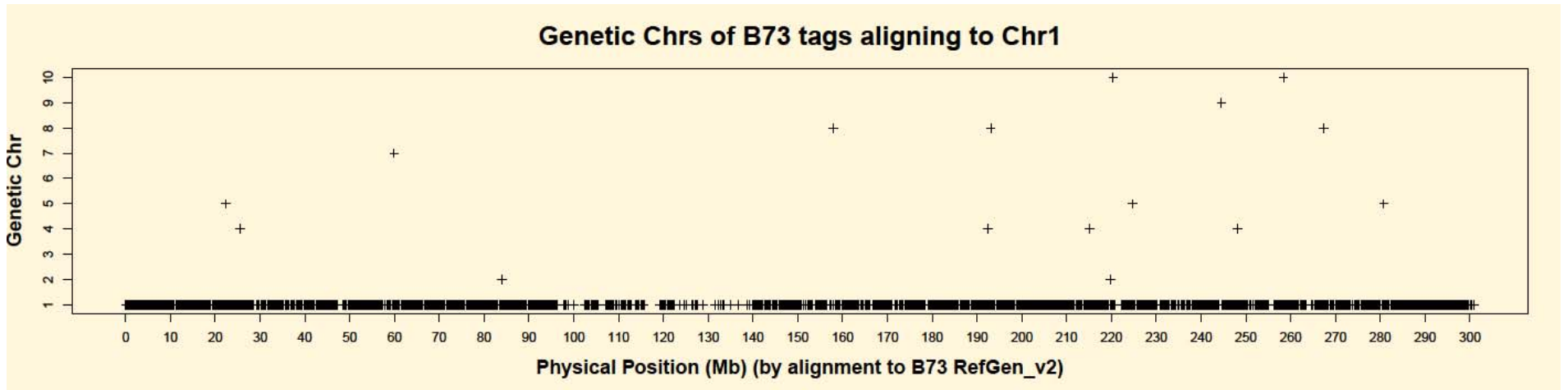
Genetically mapping individual GBS alleles in IBM

Min # Successes	<i>p</i>-value	max Recomb.	Total # GBS tags mapped	# B73 tags mapped	# Mo17 tags mapped
10	<10⁻³	<5%	485,860	266,192	219,668
20	<10⁻⁶	<5%	235,531	123,094	112,437
30	<10⁻⁷	<5%	140,713	73,829	66,884

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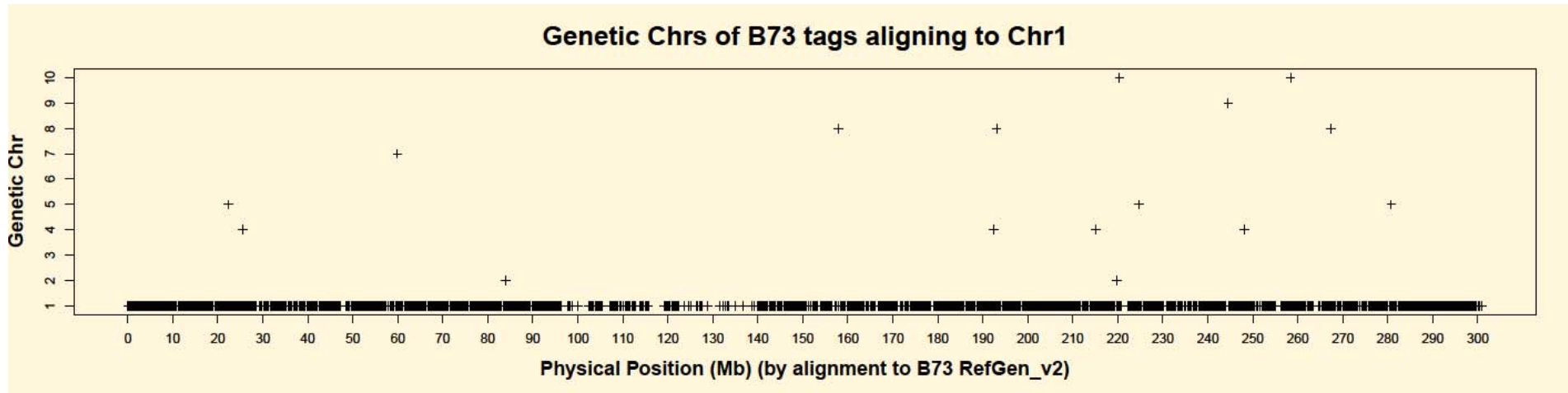
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20	$<10^{-6}$	$<5\%$	235,531	123,094	112,437
30	$<10^{-7}$	$<5\%$	140,713	73,829	66,884

B73 reference genome highly accurate for B73...



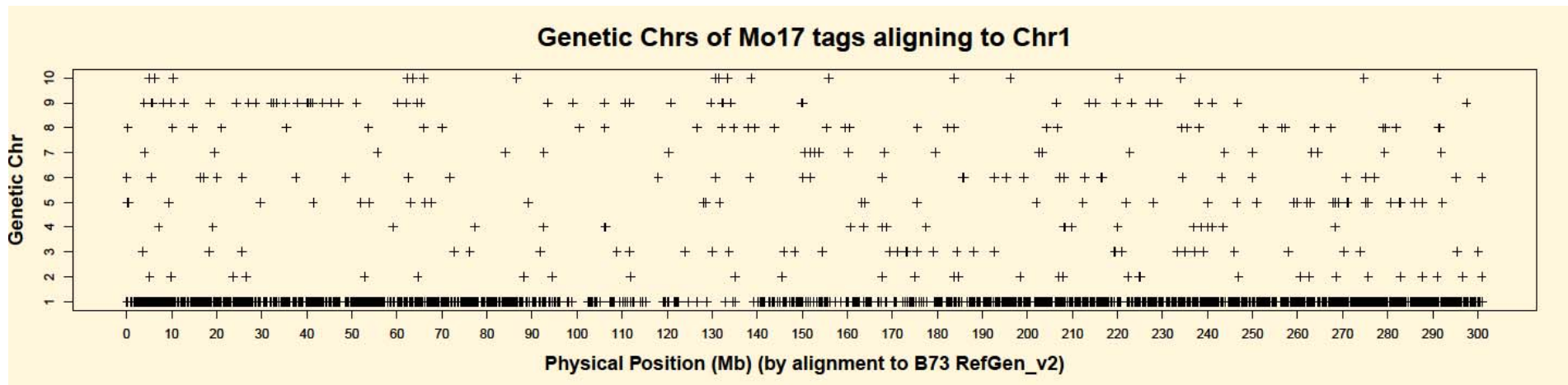
- 0.4% of B73 tags genetically map to different chromosome than they align to

B73 reference genome highly accurate for B73...



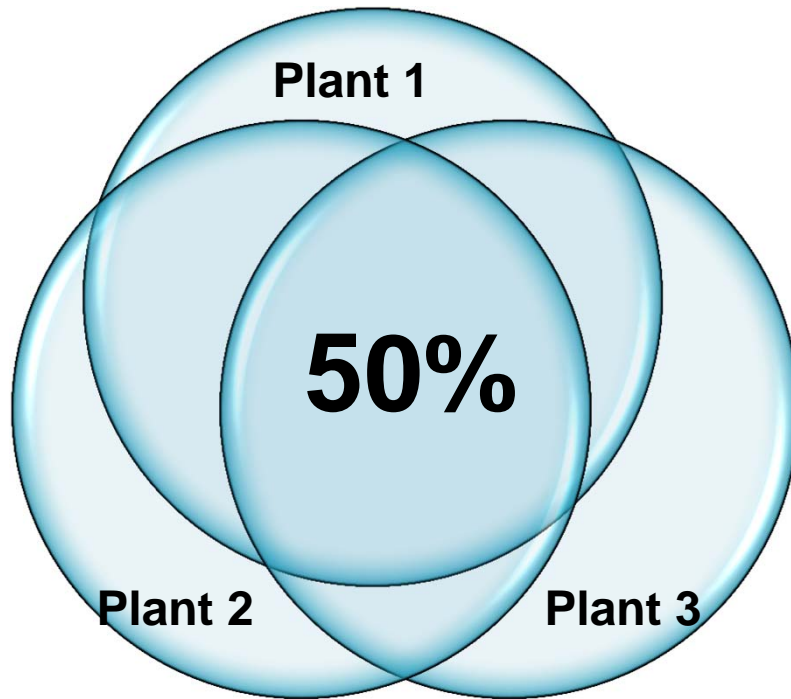
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...but far less so for other maize lines

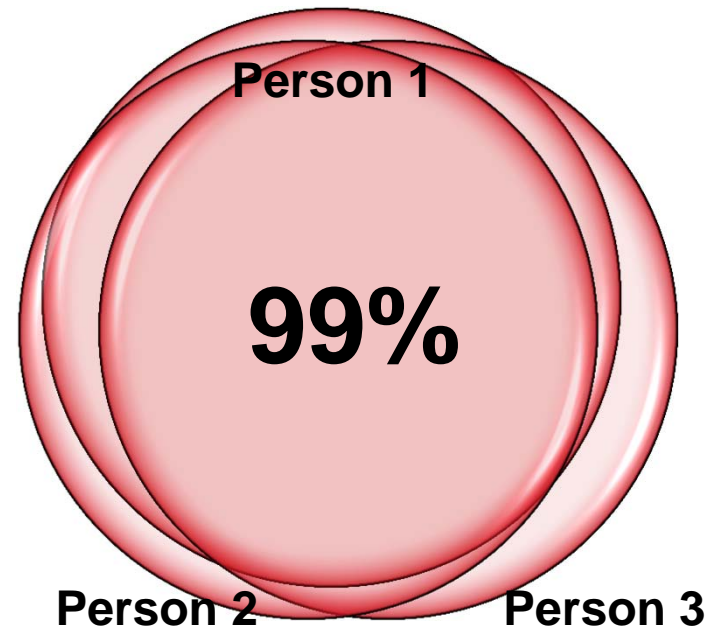


- 9.3% of Mo17 tags genetically map to different chromosome than they align to

Only 50% of the maize genome is shared between two varieties



Maize



Humans

Fu & Dooner 2002, Morgante et al. 2005, Brunner et al 2005
Numerous PAVs and CNVs - Springer, Lai, Schnable in 2010

Some chunks of the B73 reference genome are in the wrong place

Physical Chr	Start (Mb)	End (Mb)	Genetic Chr	Approx. Genetic Location (Mb)	# Tags
10	139.3	139.8	2	16.5–16.8	49
9	102.5	106.9	9	15–32	49
7	150.1	161.8	5	192–214	13
10	0.2	0.4	4	83–151	12
8	48.4	50	2	61–127	12
10	0.07	0.2	7	47–100	9
2	231.2	231.2	7	18–26	8
3	228.1	230.5	5	194–212	6

The maize B73 reference genome: room for improvement?

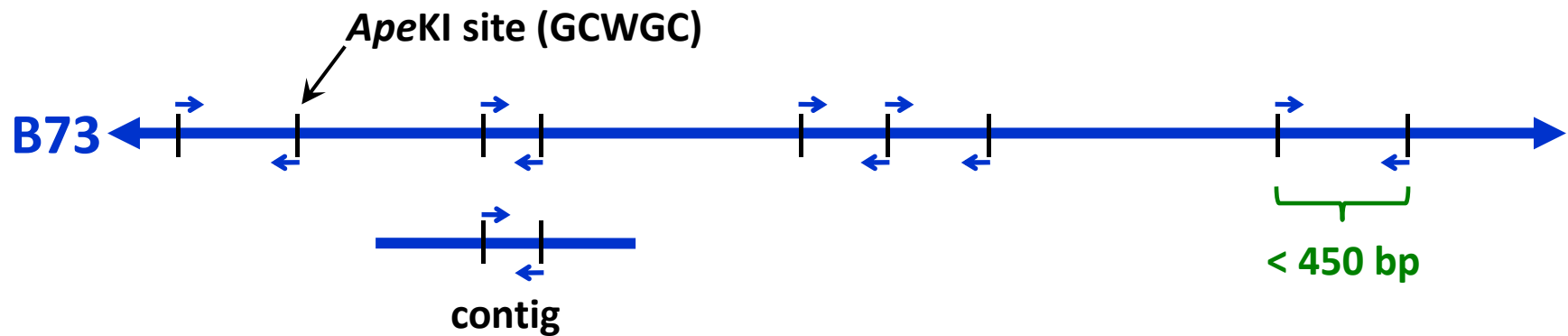
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Mapping Chr0 and *de novo* contigs via GBS

- The sequences of Chr0 contigs are known
 - so we know which *ApeKI* GBS tags are present
- *De novo* contigs constructed from 454 whole genome sequencing
 - by collaborators at CSHL (Ware *et al.*)
 - can predict *ApeKI* GBS tags from these
- Created a pipeline to genetically map novel contigs using linkage populations
- Used IBM GBS data for proof of concept

Adjacent tags on a contig or *de novo* contig can be merged into haplotypes

(→) 64-base sequence tag

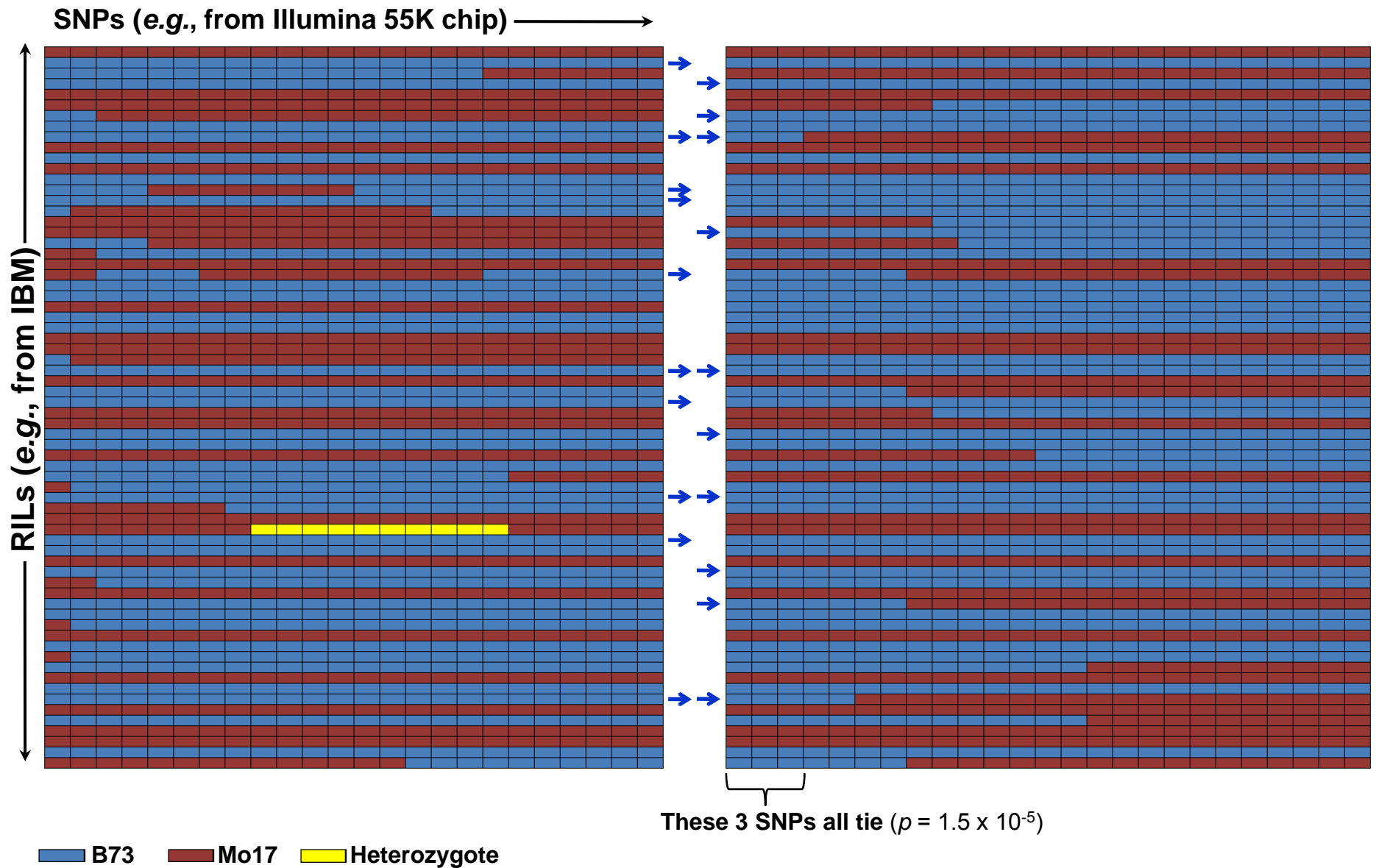


de novo (e.g., from 454 or Illumina sequence)

Novel? (not included in B73 RefGen_v2)

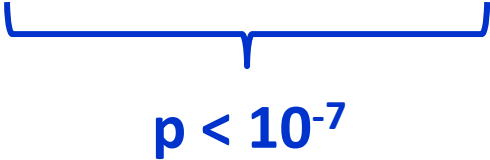
Genetically mapping GBS *haplotypes*

(→) 64-base sequence tag (GBS coverage ~0.4x)



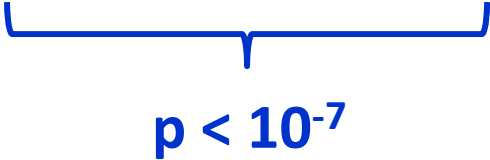
Genetically mapping contigs via GBS

<u>Contigs</u>	<u>Total #</u>	<u>Source</u>	<u># contigs genetically mapped</u>	
			<u>novel</u>	<u>non-novel</u>
Chr0	17	B73 RefGen_v2	8	---
B73 454 (k96)	3,964,387	CSHL	3,408	36,041
FLcDNA	61,477	CSHL	407	10,776

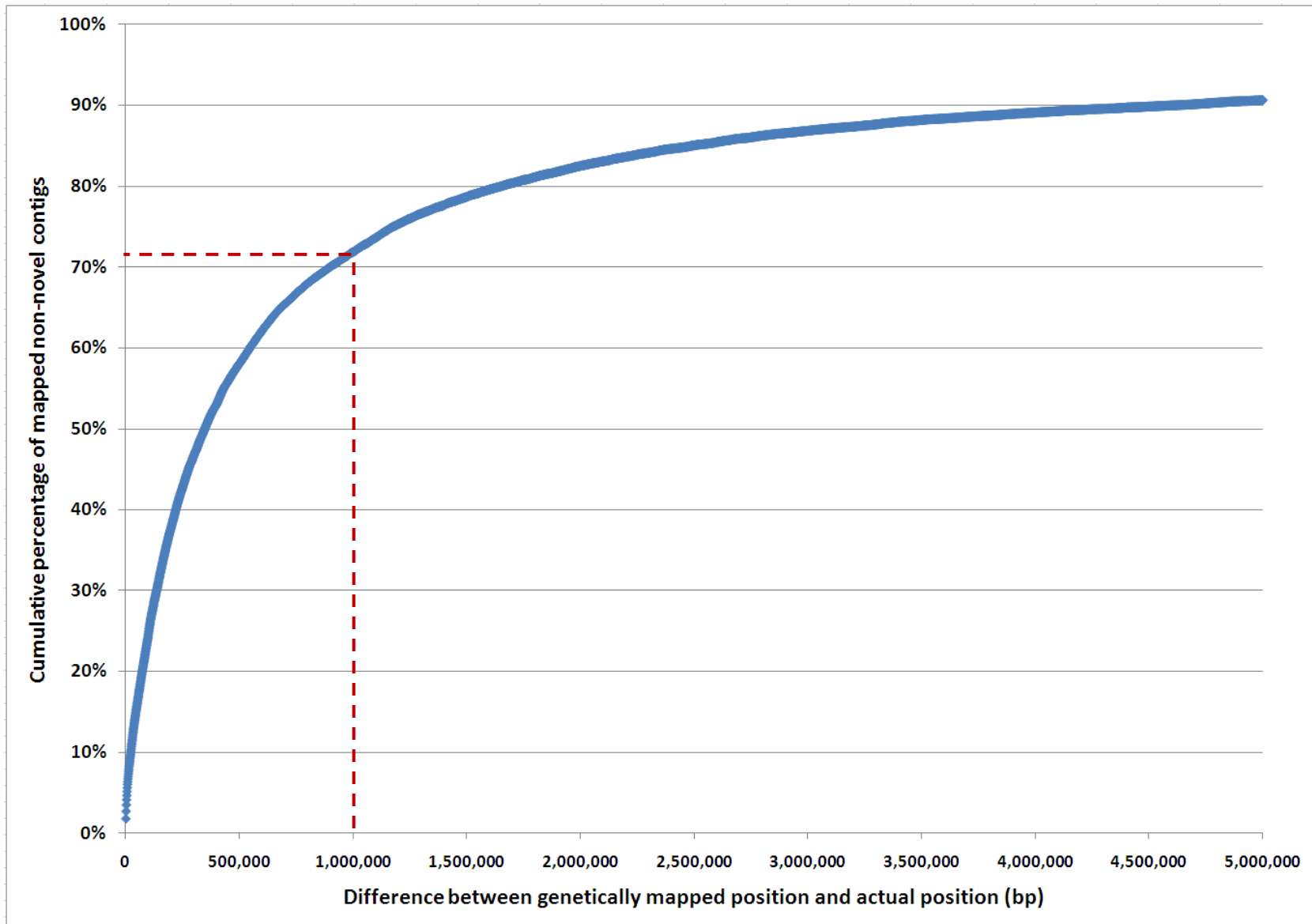

p < 10⁻⁷

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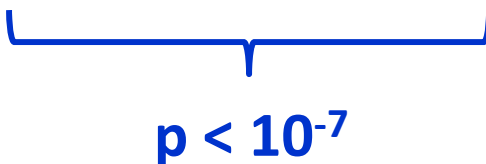

 $p < 10^{-7}$

>70% contigs genetically map to within 1 Mb of true position



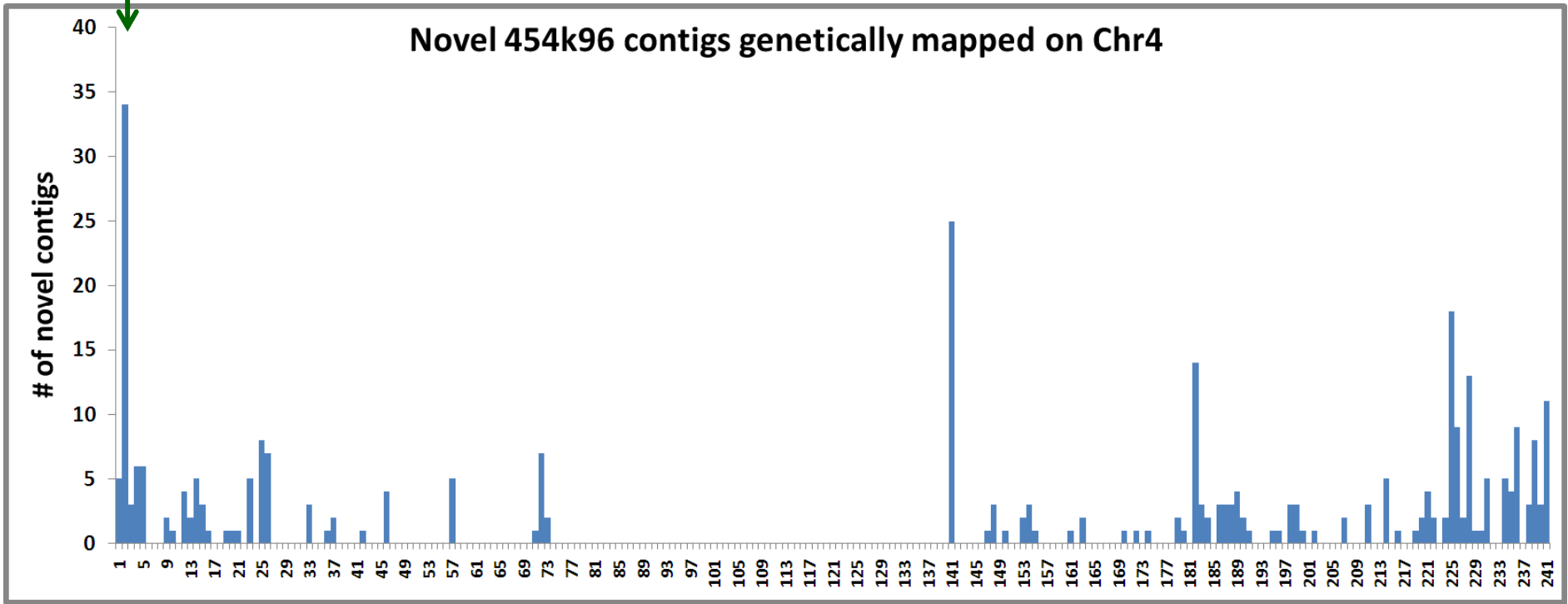
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 $p < 10^{-7}$

Some regions of reference genome are missing large chunks

Telomere of Chr4 is a prime target for future improvement



Conclusions – Improving the genome

- GBS data from a mapping population where one of the parents is the reference genome can help improve that reference genome
- Can help place:
 - unanchored contigs (chromosome 0)
 - contigs/BACS that have been misplaced (wrong chromosome)
 - novel contigs from *de novo* sequencing (missing from the reference)
- These improvements incorporated into B73 RefGenV3
- Can uncover major structural differences between lines

This coming year – Improving the genome

- **Add in GBS data from NAM for much higher resolution**
 - **Currently constructing a GBS framework map of NAM**
 - **Anchor as many novel genes & contigs as possible**
- **Use GBS SNP calls in NAM plus >10,000 additional maize lines and map tags by LD (association mapping)**
 - **Further improve genetic mapping resolution?**
 - **Preliminary results: Median resolution = 90Kb**

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