

Perl for Biologists

Session 11

May 13, 2015

Object Oriented Programming and BioPERL (2)

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Session 10 Exercises

Exercise 1. Translate all DNA sequences in a Fasta file

```
#!/usr/local/bin/perl
use strict;
use warnings;

use Bio::SeqIO;

my $in  = Bio::SeqIO->new(-file => "/home/jarekp/perl_10/yeast_orf.fasta" ,
                        -format => 'Fasta');
my $out = Bio::SeqIO->new(-file => ">yeast_pep.fasta" ,
                        -format => 'Fasta');

while ( my $seqobj = $in->next_seq() )
{
    my $proteinSeqObj = $seqobj->translate();
    $proteinSeqObj->display_id($seqobj->display_id . "_pep");
    $proteinSeqObj->desc("");
    $out->write_seq($proteinSeqObj);
};
```

Session 10 Exercises

Exercise 2. Make a fasta file with 10 random sequences

```
#!/usr/local/bin/perl
use strict;
use warnings;

use String::Random;
use Bio::SeqIO;

my $out = Bio::SeqIO->new(-file => ">random_dna.fasta" ,
                        -format => 'Fasta');

my $RandomSeq = String::Random->new();

for (my $i=0; $i<10; $i++)
{
    my $seqstr= $RandomSeq->randregex('[ACGT]{1000}');

    my $seqObject = Bio::Seq->new (-seq => $seqstr,
                                  -display_id => "seq$i",
                                  -alphabet => "dna");

    $out->write_seq($seqObject);
}
```

Review of Session 10

Bio::Seq object

A Constructor:

```
my $seqObject = Bio::Seq->new (-seq => "AAAACCCCTTGGGAAGC",  
                                -display_id => "myseq1",  
                                -desc => "This is an example.",  
                                -alphabet => "dna");
```

Methods

```
$seqObject -> revcom() -> translate(-frame=>0);
```

Alternative ways to create the sequence objects

1. From network database (e.g. NCBI Genbank)

```
use Bio::Perl;
$db = Bio::DB::GenBank->new();
$seqobj = $db->get_seq_by_acc('X78121');
```

2. From file

```
use Bio::SeqIO;
$in = Bio::SeqIO->new(-file => "inputfile.fasta" ,
                     -format => 'Fasta');
while ( my $seqobj = $in->next_seq() )
{
    ...
}
```

Other properties of Bio::Seq object

GenBank File Format

```
LOCUS      NC_000913      4639675 bp      DNA      circular BCT 04-MAR-2013
DEFINITION Escherichia coli str. K-12 substr. MG1655, complete genome.
ACCESSION  NC_000913
VERSION    NC_000913.2  GI:49175990
DBLINK     Project: 57779
           BioProject: PRJNA57779

KEYWORDS   .
SOURCE     Escherichia coli str. K-12 substr. MG1655
  ORGANISM Escherichia coli str. K-12 substr. MG1655
           Bacteria; Proteobacteria; Gammaproteobacteria; Enterobacteriales;
           Enterobacteriaceae; Escherichia.

FEATURES   Location/Qualifiers
    source  1..4639675
            /organism="Escherichia coli str. K-12 substr. MG1655"
            /mol_type="genomic DNA"
            /strain="K-12"
            /sub_strain="MG1655"
            /db_xref="taxon:511145"
    gene    190..255
            /gene="thrL"
            /locus_tag="b0001"
            /gene_synonym="ECK0001; JW4367"
            /db_xref="EcoGene:EG11277"
            /db_xref="GeneID:944742"
    CDS     190..255
            /gene="thrL"
            /locus_tag="b0001"
            /gene_synonym="ECK0001; JW4367"
            /function="leader; Amino acid biosynthesis: Threonine"
            /function="1.5.1.8 metabolism; building block
            biosynthesis; amino acids; threonine"
            /GO_process="GO:0009088 - threonine biosynthetic process"
```

Other Bio::Seq properties: Seq Features

GFF3 File Format

Chr1	TAIR10	chromosome	1	30427671	.	.	.	ID=Chr1;Name=Chr1
Chr1	TAIR10	gene	3631	5899	.	+	.	ID=AT1G01010;Note=prote
Chr1	TAIR10	mRNA	3631	5899	.	+	.	ID=AT1G01010.1;Parent=AT
Chr1	TAIR10	protein	3760	5630	.	+	.	ID=AT1G01010.1-Protein;N
Chr1	TAIR10	exon	3631	3913	.	+	.	Parent=AT1G01010.1
Chr1	TAIR10	five_prime_UTR		3631	3759	.	+	Parent=AT1G
Chr1	TAIR10	CDS	3760	3913	.	+	0	Parent=AT1G01010.1,AT1G
Chr1	TAIR10	exon	3996	4276	.	+	.	Parent=AT1G01010.1
Chr1	TAIR10	CDS	3996	4276	.	+	2	Parent=AT1G01010.1,AT1G
Chr1	TAIR10	exon	4486	4605	.	+	.	Parent=AT1G01010.1
Chr1	TAIR10	CDS	4486	4605	.	+	0	Parent=AT1G01010.1,AT1G
Chr1	TAIR10	exon	4706	5095	.	+	.	Parent=AT1G01010.1
Chr1	TAIR10	CDS	4706	5095	.	+	0	Parent=AT1G01010.1,AT1G
Chr1	TAIR10	exon	5174	5326	.	+	.	Parent=AT1G01010.1
Chr1	TAIR10	CDS	5174	5326	.	+	0	Parent=AT1G01010.1,AT1G
Chr1	TAIR10	exon	5439	5899	.	+	.	Parent=AT1G01010.1
Chr1	TAIR10	CDS	5439	5630	.	+	0	Parent=AT1G01010.1,AT1G
Chr1	TAIR10	three_prime_UTR		5631	5899	.	+	Parent=AT1G
Chr1	TAIR10	gene	5928	8737	.	-	.	ID=AT1G01020;Note=prote
Chr1	TAIR10	mRNA	5928	8737	.	-	.	ID=AT1G01020.1;Parent=AT
Chr1	TAIR10	protein	6915	8666	.	-	.	ID=AT1G01020.1-Protein;N
Chr1	TAIR10	five_prime_UTR		8667	8737	.	-	Parent=AT1G
Chr1	TAIR10	CDS	8571	8666	.	-	0	Parent=AT1G01020.1,AT1G
Chr1	TAIR10	exon	8571	8737	.	-	.	Parent=AT1G01020.1
Chr1	TAIR10	CDS	8417	8464	.	-	0	Parent=AT1G01020.1,AT1G
Chr1	TAIR10	exon	8417	8464	.	-	.	Parent=AT1G01020.1
Chr1	TAIR10	CDS	8236	8325	.	-	0	Parent=AT1G01020.1,AT1G

Other Bio::Seq properties: Seq Features

GFF3 File Format

Chr1	TAIR10	chromosome	1	30427671	.	.	.	ID=Chr1;Name=Chr1
Chr1	TAIR10	gene	3631	5899	.	+	.	ID=AT1G01010;Note=prote
Chr1	TAIR10	mRNA	3631	5899	.	+	.	ID=AT1G01010.1;Parent=AT
Chr1								AT1G01010.1-Protein;N
Chr1								AT1G01010.1
								Parent=AT1G
								AT1G01010.1,AT1G
								AT1G01010.1
								AT1G01010.1,AT1G
								AT1G01010.1
								AT1G01010.1,AT1G
								AT1G01010.1
								AT1G01010.1,AT1G
								AT1G01010.1
								AT1G01010.1,AT1G
								Parent=AT1G
								Note=prote
								1;Parent=AT
								AT1G01010.1-Protein;N
								Parent=AT1G
Chr1	TAIR10	CDS	8571	8666	.	-	0	Parent=AT1G01020.1,AT1G
Chr1	TAIR10	exon	8571	8737	.	-	.	Parent=AT1G01020.1
Chr1	TAIR10	CDS	8417	8464	.	-	0	Parent=AT1G01020.1,AT1G
Chr1	TAIR10	exon	8417	8464	.	-	.	Parent=AT1G01020.1
Chr1	TAIR10	CDS	8236	8325	.	-	0	Parent=AT1G01020.1,AT1G

```

open (IN, "tair10.gff3") || die "Can not open GFF3 file!\n";

while (<IN>)
{
    my @data = split "\t";
    ...
}

```


Retrieve seq features from a Bio:Seq object constructed from NCBI Genbank

script1.pl

```
#!/usr/local/bin/perl
use strict;
use warnings;
use Bio::Perl;

my $db = Bio::DB::GenBank->new();
my $seqobj = $db->get_Seq_by_acc('NC_000913');

$,="\t";
my $count = 0;
for my $feat_object ($seqobj->get_SeqFeatures) {
    if ($feat_object->primary_tag eq "gene") {
        $count ++;
        print $feat_object->get_tag_values('gene'),
              $feat_object->start(),
              $feat_object->end(),
              $feat_object->strand(), "\n";
    }
}
print "Total number of genes: $count\n";
```

Run Sequence Analysis tools

1. Using BioPERL wrapper: Bio::Tools::Run

ClusterW

MUSCLE

BLAST

...

Primer3

...

2. Using system calling

```
system ("primer3_core < inputFile");
```

Or

```
my $stdout = ` primer3_core < inputFile `;
```

Using Bio::Tools::Run::Primer3

script2.pl

```
#!/usr/local/bin/perl
use strict;
use warnings;
use Bio::DB::GenBank;
use Bio::Tools::Run::Primer3;

my $db = Bio::DB::GenBank->new();
my $seqobj = $db->get_seq_by_acc('NM_001126114');

my $primer3 = Bio::Tools::Run::Primer3->new(
    -seq => $seqobj,
    -outfile => "temp.out",
    -path => "/programs/primer3-2.3.5/src/primer3_core");

$primer3->add_targets(
    "PRIMER_MIN_TM"=>56,
    "PRIMER_MAX_TM"=>90,
    "PRIMER_MIN_SIZE"=>18,
    "PRIMER_MAX_SIZE"=>21);

my $results = $primer3->run;
print "There were ", $results->number_of_results, " primers\n";
```

Bio::Tools::Run::Primer3 does not work with latest version of Primer3

Parameter name is changed after Primer3 2.0

Boulder data interchange format

```
SEQUENCE_ID=example
SEQUENCE=GTAGTCAGTAGACNATGACNACTGACGATGCAGACNAC
ACACACACACACAGCACACAGGTATTAGTGGGCCATTTCGATCCCGACC
CAAATCGATAGCTACCATGACG
SEQUENCE_TARGET=37,21
PRIMER_TASK=pick_detection_primers
PRIMER_PICK_LEFT_PRIMER=1
PRIMER_PICK_INTERNAL_OLIGO=1
PRIMER_PICK_RIGHT_PRIMER=1
PRIMER_OPT_SIZE=18
PRIMER_MIN_SIZE=15
PRIMER_MAX_SIZE=21
PRIMER_MAX_NS_ACCEPTED=1
PRIMER_PRODUCT_SIZE_RANGE=75-100
P3_FILE_FLAG=1
SEQUENCE_INTERNAL_EXCLUDED_REGION=37,21
PRIMER_EXPLAIN_FLAG=1
=
```

Tag name is changed to
SEQUENCE_TEMPLATE
In latest version.

Using Bio::Tools::Run::Primer3

script3.pl

```
#!/usr/local/bin/perl
use strict;
use warnings;
use Bio::DB::GenBank;
my $PRIMER_MIN_TM=56;
my $PRIMER_MAX_TM=90;
my $PRIMER_MIN_SIZE=15;
my $PRIMER_MAX_SIZE=21;

my $db = Bio::DB::GenBank->new();
my $seqobj = $db->get_seq_by_acc('NM_001126114');
my $seqid = $seqobj->display_id();
my $seqstr = $seqobj->seq();

open OUT, ">temp.input";
print OUT <<EOF;
SEQUENCE_ID=$seqid
SEQUENCE_TEMPLATE=$seqstr;
PRIMER_MIN_TM=$PRIMER_MIN_TM
PRIMER_MAX_TM=$PRIMER_MAX_TM
PRIMER_MIN_SIZE=$PRIMER_MIN_SIZE
PRIMER_MAX_SIZE=$PRIMER_MAX_SIZE
PRIMER_LIBERAL_BASE=1
=
EOF
close OUT;
system "/programs/primer3-2.3.5/src/primer3_core -output=temp.output temp.input";
```

Parsing results from analysis software

Output from codeml

```
.....
.....
Model 1: NearlyNeutral (2 categories)

TREE # 1: ((3, 4), 2, (1, 5)); MP score: 0
lnL(ntime: 7 np: 10): -548.665307 +0.000000
  6..7  7..3  7..4  6..2  6..8  8..1  8..5
  0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 1.98425 0.60769 0.54695
Note: Branch length is defined as number of nucleotide substitutions per codon (not per neucleotide site).

tree length = 0.00002

((3: 0.000005, 4: 0.000005): 0.000000, 2: 0.000005, (1: 0.000005, 5: 0.000005): 0.000000);

((CT18: 0.000005, Ty2: 0.000005): 0.000000, ch: 0.000005, (ATCC9150: 0.000005, LT2: 0.000005): 0.000000);

Detailed output identifying parameters

kappa (ts/tv) = 1.98425

dN/dS for site classes (K=2)

p: 0.60769 0.39231
w: 0.54695 1.00000

dN & dS for each branch

branch      t      S      N  dN/dS   dN     dS  S*dS  N*dN
6..7    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0
7..3    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0
7..4    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0
6..2    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0
6..8    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0
8..1    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0
8..5    0.000  116.9  291.1  0.7247  0.0000  0.0000  0.0  0.0

.....
.....
```

.....

 Model 1: NearlyNeutral (2 categories)

TR

PAML Parser

```
use Bio::Tools::Phylo::PAML;
my $parser = Bio::Tools::Phylo::PAML->new(
    -file => "./output.mlc",
    -dir  => "./",
    -ctlf => "./codeml.ctl");

while(my $result = $parser->next_result) {
    # do something with the results from this dataset
    ...
}
```

```
6..2
6..8 0.000 116.9 291.1 0.7247 0.0000 0.0000 0.0 0.0
8..1 0.000 116.9 291.1 0.7247 0.0000 0.0000 0.0 0.0
8..5 0.000 116.9 291.1 0.7247 0.0000 0.0000 0.0 0.0
```

.....

Parse Blast Results

```
blastall -p blastp -i rice.fasta -d TAIR7_pep_db -o blastresults
```

Note:

Most new software starts to provide machine readable output files, e.g. NCBI BLAST

-m 7 : XML (used by Blast2GO, et al.)

-m 8 : tab delimited text file (used by OrthoMCL, et al.)

BLAST Results

Query= Os01g01010.1
(702 letters)

Database: TAIR7_pep_20070320
31,921 sequences; 13,036,889 total letters

Searching.....done

Sequences producing significant alignments:			Score (bits)	E Value
AT2G43490.1	Symbols:	RabGAP/TBC domain-containing protein ...	621	0.0
AT3G59570.1	Symbols:	RabGAP/TBC domain-containing protein ...	608	0.0
AT5G54780.1	Symbols:	RAB GTPase activator chr5:22265922-2...	184	4e-051
AT4G27100.2	Symbols:	RAB GTPase activator chr4:13595851-1...	183	6e-051
AT4G27100.1	Symbols:	RAB GTPase activator chr4:13595851-1...	182	9e-051
AT2G20440.1	Symbols:	RabGAP/TBC domain-containing protein ...	175	4e-048
AT4G28550.1	Symbols:	RabGAP/TBC domain-containing protein ...	170	2e-046
AT5G41940.1	Symbols:	RabGAP/TBC domain-containing protein ...	136	6e-034
AT5G53570.1	Symbols:	RabGAP/TBC domain-containing protein ...	134	4e-033
AT5G24390.1	Symbols:	RabGAP/TBC domain-containing protein ...	130	5e-032
.....	.			
.....	.			

BLAST Results

Query Object

Query name

Query length

Query= **Os01g01010.1**
(**702** letters)

Database: TAIR7_pep_20070320
31,921 sequences; 13,036,889 total letters

Searching.....done

Sequences producing significant alignments:				Score (bits)	E Value
AT2G43490.1	Symbols:	RabGAP/TBC domain-containing protein	...	621	0.0
AT3G59570.1	Symbols:	RabGAP/TBC domain-containing protein	...	608	0.0
AT5G54780.1	Symbols:	RAB GTPase activator	chr5:22265922-2...	184	4e-051
AT4G27100.2	Symbols:	RAB GTPase activator	chr4:13595851-1...	183	6e-051
AT4G27100.1	Symbols:	RAB GTPase activator	chr4:13595851-1...	182	9e-051
AT2G20440.1	Symbols:	RabGAP/TBC domain-containing protein	...	175	4e-048
AT4G28550.1	Symbols:	RabGAP/TBC domain-containing protein	...	170	2e-046
AT5G41940.1	Symbols:	RabGAP/TBC domain-containing protein	...	136	6e-034
AT5G53570.1	Symbols:	RabGAP/TBC domain-containing protein	...	134	4e-033
AT5G24390.1	Symbols:	RabGAP/TBC domain-containing protein	...	130	5e-032

..... *

..... *

>AT4G27100.2 RAB GTPase activator
Length = 433

Score = 183 bits (464), Expect = 6e-051, Method: Compositional matrix adjust.
Identities = 91/188 (48%), Positives = 122/188 (64%), Gaps = 10/188 (5%)

Query: 370 GTKSNSVVASKD-----RVSEWLWTLHRIVDVVRTDSHLDFYGESRNMARMSDIL 420
GT SN V K+ ++ +WL TLH+I +DV RTD L FY + N++++ DIL

Sbjct: 144 GTNSNGSVFFKELTSRGPLDKKIIQWLLTLHQIGLDVNRTDRALVFYEKKENLSKLWDIL 203

Query: 421 AVYAWVDPSTGYCQGMSDLSPFVVLIEDDADAFWCFEMLLRRMRENFQMEG-PTGVMKQ 479
+VYAW+D GYCQGMSDL SP ++L ED+ADAFWCFE L+RR+R NF+ G GV Q

Sbjct: 204 SVYAWIDNDVGYCQGMSDLCSPMIILLEDEADAFWCFERLMRRLRGNFRSTGRSVGVEAQ 263

Query: 480 LQALWKIMEITDVELFEHLSTIGAESLHFAFRMLLVLFRRRELSFEESLSMWEMMWAAADFN 539
L L I ++ D +L +HL +G FA RML+V FRRE SF +SL +WEMMWA +++

Sbjct: 264 LTHLSSITQVDPKLHQHLDKLGGDYLF AIRMLMVQFRREFSFCDSLWLWEMMWALEYD 323

Query: 540 EDVILHLE 547
D+ E

Sbjct: 324 PDLFYVYE 331

Score = 65.1 bits (157), Expect = 5e-011, Method: Compositional matrix adjust.
Identities = 42/96 (43%), Positives = 54/96 (56%), Gaps = 3/96 (3%)

Query: 55 VKGSKMLKPEKWHTCFDNDGKV-IGFRKALKFIVLGGVDPTIRAEVWEFLLGCYALSSTS 113
+K K L KW F +G + IG K L+ I GG+ P+IR EVWEFLLGCY ST

Sbjct: 29 IKPGKTL SVRKWQAVFVQEGSLHIG--KTLRRIRRGGIHPSIRGEVWEFLLGCYDPMSTF 86

Query: 114 EYRRKLRAVRREKYQILVRQCQSMHPSIGTGELAYA 149
E R ++R RR +Y +C+ M P IG+G A

Sbjct: 87 EEREQIRQRRRLQYASWKEECKQMFPVIGSGRFTTA 122

Hit

HSP 1

HSP 2

>AT4G27100.2 RAB GTPase activator
Length = 433

Score = 183 bits (464), Expect = 6e-051, Method:
Identities = 91/188 (48%), Positives = 122/188 (64%)

Query: 370 GTKSNSVVASKD-----RVSEWLWTLHRIV
GT SN V K+ ++ +WL TLH+I

Sbjct: 144 GTNSNGSVFFKELTSRGPLDKKIIQWLLTLHQIC

Query: 421 AVYAWVDPSTGYCQGMSDLSPFVVLIEDDADAF
+VYAW+D GYCQGMSDL SP ++L ED+ADAF

Sbjct: 204 SVYAWIDNDVGYCQGMSDLCSPMIILLEDEADAF

Query: 480 LQALWKIMEITDVELFEHLSTIGAESLHFAFRML
L L I ++ D +L +HL +G FA RML

Sbjct: 264 LTHLSSITQVDPKLHQHLDKLGGDYLF AIRML

Query: 540 EDVILHLE 547
D+ E

Sbjct: 324 PDLFYVYE 331

Score = 65.1 bits (157), Expect = 5e-011, Method: Compositional matrix adjust.
Identities = 42/96 (43%), Positives = 54/96 (56%), Gaps = 3/96 (3%)

Query: 55 VKGSKMLKPEKWHTCFDNDGKV-IGFRKALKFIVLGGVDPTIRAEVWEFLLGCYALSSTS 113
+K K L KW F +G + IG K L+ I GG+ P+IR EVWEFLLGCY ST

Sbjct: 29 IKPGKTL SVRKWQAVFVQEGSLHIG--KTLRRIRRGGIHPSIRGEVWEFLLGCYDPMSTF 86

Query: 114 EYRRKLRAVRREKYQILVRQCQSMHPSIGTGELAYA 149
E R ++R RR +Y +C+ M P IG+G A

Sbjct: 87 EEREQIRQRRRLQYASWKEECKQMFPVIGSGRFTTA 122

Each hit object:

Hit name

Hit length

Hsps

adjust.

Hit

Each HSP object:

Query: start - end - strand

Hit: start - end - strand

Bit score

E-value

Identities

Positives

Alignment length

Gaps

Query sequence

Hit sequence

HSP 1

HSP 2

BLAST Parser

script4.pl

```
#!/usr/local/bin/perl
use Bio::SearchIO;
($infile, $outfile) = @ARGV;

open OUT, ">$outfile";
$,="\t";

$searchio = Bio::SearchIO->new(-format => 'blast',
                              -file   => $infile);
while ($result = $searchio->next_result)
{
    # Get info about the entire report
    $query_name = $result->query_name;
    $query_length = $result->query_length;

    # get info about the first hit
    while ($hit = $result->next_hit)
    {
        $hit_name = $hit->name;
        $hit_length = $hit->length;

        # get info about the first hsp of the first hit
        while ($hsp = $hit->next_hsp)
        {
            $rank = $hsp->rank;
            $num_conserved = $hsp->num_conserved ;
        }
    }
}
```

Loop1: Query

Loop2: Hit

Loop3: HSP

BLAST Parser

```
while ($hit = $result->next_hit)
{
    $hit_name = $hit->name;
    $hit_length = $hit->length;

    # get info about the first hsp of the first hit
    while ($hsp = $hit->next_hsp){
        $rank = $hsp->rank;
        $num_conserved = $hsp->num_conserved ;
        $num_identical= $hsp->num_identical ;
        $hsp_length= $hsp->hsp_length ;
        $bits= $hsp->bits ;
        $evalue = $hsp->evalue ;
        $hsp_qstart = $hsp->query->start;
        $hsp_qend = $hsp->query->end;
        $query_strand = $hsp->query->strand;
        $hsp_hstart = $hsp->hit->start;
        $hsp_hend = $hsp->hit->end;
        $hit_strand = $hsp->hit->strand;
        $query_string = $hsp->query_string ;
        $hit_string = $hsp->hit_string ;
        $homology_string = $hsp->homology_string ;

        print OUT 1, $query_name, $hit_name, $query_length, $hit_length, $rank, $num_identical,
        $num_conserved, $hsp_length, $bits, $evalue, $hsp_qstart, $hsp_qend, $query_strand, $hsp_hstart, $hsp_hend, $hit_strand,
        $query_string, $hit_string, $homology_string, "", "";
        print OUT "\n";
    }
}
```

script4.pl

Query

Hit

HSP

Parsed results from BLAST

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V
1	1	Os01g010:AT2G4349	702	756	1	345	452	730	621		0	12	685	0	3	689	0	TAGDYIKWMCXXXXXX)SGGEGKQWSCGKA(+G+ +W C		+LQ+ VGS LVRD+ +P		
2	1	Os01g010:AT3G5957	702	720	1	334	435	687	608		0	12	685	0	3	653	0	TAGDYIKWMCXXXXXX)SAGEGKKW-----TRF+AG+ KW		+LQ+ V S LVRD+ +PC		
3	1	Os01g010:AT5G5478	702	432	1	109	157	300	184	4.00E-51	382	680	0	163	414	0	RVSEWLWTLHRIVVDV\ KVIQWLLTLHQIGLD	+V +WL TLH+I +DV RTD	L FY + N+++			
4	1	Os01g010:AT5G5478	702	432	2	40	51	95	64.7	7.00E-11	55	149	0	29	122	0	VKGSKMLKPEKWHTCF\IKPGKTL SVRKWQA	+K K L KW F +G + K L I	GG+ P+H			
5	1	Os01g010:AT4G2710	702	433	1	91	122	188	183	6.00E-51	370	547	0	144	331	0	GTKSNSVVASKD-----GTNSNGSVFFKELTS	GT SN V K+		++ +WL TLH+I +DV F		
6	1	Os01g010:AT4G2710	702	433	2	42	54	96	65.1	5.00E-11	55	149	0	29	122	0	VKGSKMLKPEKWHTCF\IKPGKTL SVRKWQA	+K K L KW F +G +IG K L+I	GG+ P-			
7	1	Os01g010:AT4G2710	702	433	3	17	30	51	37	0.035	632	680	0	365	415	0	AKNGDDDLPI--FCVAAI GKSAEGLPISVFLV\	K++ LPI F VA++L	+K++ E R +DD +K			
8	1	Os01g010:AT4G2710	702	436	1	91	122	188	182	9.00E-51	370	547	0	144	331	0	GTKSNSVVASKD-----GTNSNGSVFFKELTS	GT SN V K+		++ +WL TLH+I +DV F		
9	1	Os01g010:AT4G2710	702	436	2	42	54	96	65.1	5.00E-11	55	149	0	29	122	0	VKGSKMLKPEKWHTCF\IKPGKTL SVRKWQA	+K K L KW F +G +IG K L+I	GG+ P-			
10	1	Os01g010:AT4G2710	702	436	3	17	30	51	37	0.036	632	680	0	365	415	0	AKNGDDDLPI--FCVAAI GKSAEGLPISVFLV\	K++ LPI F VA++L	+K++ E R +DD +K			
11	1	Os01g010:AT2G2044	702	425	1	95	157	310	175	4.00E-48	377	685	0	154	409	0	VASKDRVSEWLWTLHRI TVTDERVLQWMLSL	+ +RV +W+ +LH+I +DV RTD	+L FY RI			
12	1	Os01g010:AT2G2044	702	425	2	39	51	90	80.1	9.00E-16	56	145	0	37	125	0	KGSKMLKPEKWHTCFD\RAGKTL SARRWHAA	+ K L +WH F DG + K L+I	GG+ P+H			
13	1	Os01g010:AT4G2855	702	424	1	79	112	168	170	2.00E-46	382	548	0	159	326	0	RVSEWLWTLHRIVVDV\ RVLQWMLVLSQIGL	RV +W+ L +I +DVVRTD	+L FY N AR+			
14	1	Os01g010:AT4G2855	702	424	2	39	53	92	80.9	4.00E-16	56	147	0	37	127	0	KGSKMLKPEKWHTCFD\RAGKTL SARKWHAA	+ K L KWH F DG + +L+I	GG+ P+H			
15	1	Os01g010:AT5G4194	702	549	1	60	84	122	136	6.00E-34	414	535	0	345	465	0	ARMSDILAVYAWVDP\ARLVGILEAYAVYDP	AR+ IL YA DP	GYCQGMSDLLSP ++			
16	1	Os01g010:AT5G4194	702	549	2	48	72	135	72.4	4.00E-13	45	176	0	76	205	0	IGDPCLNPSPVKGSKML\IGSPW---SLRRRKRV	IG P S + +L+P++W+ F +G++	G K			
17	1	Os01g010:AT5G5357	702	550	1	59	86	122	134	4.00E-33	414	535	0	338	458	0	ARMSDILAVYAWVDP\ARLVAILAYAMYDF	AR+ IL YA DP	GYCQGMSDLLSP ++			
18	1	Os01g010:AT5G5357	702	550	2	37	53	81	72.8	3.00E-13	61	137	0	96	173	0	LKPEKWHTCFDNDGKV-LTPHQWRSFLTPEG	L P +W+ F +GK+ +GF	LK+ G VDP+			
19	1	Os01g010:AT5G2439	702	528	1	56	87	124	130	5.00E-32	412	535	0	315	437	0	NMARMSDILAVYAWV\HAARLVAVLEAYAL	+AR+ +L YA DP	GYCQGMSDLLSP +			
20	1	Os01g010:AT5G2439	702	528	2	38	52	96	65.9	4.00E-11	47	138	0	46	141	0	DPCLNPSP---VKGSKMLDPHRLKSPWSRRKG	DP SP KG K L +W CF	+G++ G L			
21	1	Os01g010:AT3G4935	702	539	1	57	88	133	127	9.00E-31	403	535	0	320	450	0	HLDFYGESRNMARMSD\HLEPY-KIFQAARLV	HL+Y+ AR+ +L YA DP	GYCQGMSSI			
22	1	Os01g010:AT3G4935	702	539	2	30	46	76	60.8	2.00E-09	64	138	0	72	147	0	EKWHTCFDNDGKVI-GFQQWKRFFTPDGRLF	++W F DG++ G	LK+ G++P+IR EV			
23	1	Os01g010:AT5G5259	702	338	1	57	90	153	112	8.00E-27	396	546	0	107	258	0	DVVRTDSHLDFYGESRN DVVRTDRAFEYEGE	DVVRTD ++Y N+	M DIL Y+++ G'			
24	1	Os01g010:AT5G5259	702	338	2	34	50	93	59.7	2.00E-09	70	159	0	19	111	0	FDNDGKVIGFRKALKFIVLDSEGRVVESKALRE	D++G+V+ + +	GG++ +R EVW FLL			
25	1	Os01g010:AT4G1373	702	408	1	49	91	177	77.4	6.00E-15	389	546	0	227	403	0	TLHRIVVDVVRTDSHLDFVLEQIERDVMRTHPI	L+I DV+RT +F+	+A+ ++IL+AA			
26	1	Os01g010:AT4G1373	702	449	1	47	87	164	76.6	1.00E-14	389	533	0	227	390	0	TLHRIVVDVVRTDSHLDFVLEQIERDVMRTHPI	L+I DV+RT +F+	+A+ ++IL+AA			
27	1	Os01g010:AT1G0483	702	448	1	56	90	186	72.8	2.00E-13	389	555	0	223	398	0	TLHRIVVDVVRTDSHLDFTIEQIDRDVKRTHPD	T++I DV RT +F+	+AR M +IL V+Z			
28	1	Os01g010:AT2G3071	702	440	1	45	79	169	71.6	5.00E-13	390	535	0	201	368	0	LHRIVVDVVRTDSHLDFLRQIAVDCPRTVPD	L+I VD RT +F+++	+ IL +A P++G			
29	1	Os01g010:AT2G1924	702	840	1	32	51	109	49.3	7.00E-06	444	537	0	248	356	0	VVLYED--DADAFWCFE\IVLSEKFMEHDAYCM	+VLE +DA+ F+L+ + F M	G'			

Sequence Alignment

CLUSTAL W(1.81) multiple sequence alignment

Clustalw format

```
seq1      VANITLSTQHYRIHRSDVEPVKEKTTDKDVFAKSITAVRNSFISLSTSLSDRFSLHLQTD
seq2      VTNITLSTQHYRIHRSDVEPVKEKTEKDIFAKSITAVRNSFISLSTSLSDRFSLHQQTd
seq3      VTNITLSTQHYRIHRSDVEPVKEKTEKDIFAKSITAVRNSFISLSTSLSDRFSLHQQTd
seq4      VTKITLSPQNFRIQKQET--LKEKSTEKNSLAKSILAVKNHFIELRSKLSERFISHKNTe
seq5      VTKITLSPQNFRIQKQETTLLKEKSTEKNSLAKSILAVKNHFIELRSKLSERFISHKNTe
          *:*****.*::**::.. :*****: :**** **:* **.* ::*** ** *::
```

```
seq1      IPTTHFHRRGSASEGRAVL TSKTVKDFMLQKLNSLDIKGNA
seq2      IPTTHFHRRGSASEGRAVL TSKTVKDFMLQKLNSLDIKGNA
seq3      IPTTHFHRRGSASEGRAVL TSKTVKDFMLQKLNSLDIKGNA
seq4      SSATHFHRRGSASEGRAVL TNKVVKDFMLQTLNDIDIRGSA
seq5      SSATHFHRRGSASEGRAVL TNKVVKDFMLQTLNDIDIRGSA
          .*****.*****.******.***.*.*.*
```

Phylip format

	5	100					
seq1	VANITLSTQH	YRIHRSDVEP	VKEKTTDKDV	FAKSITAVRN	SFISLSTSL	DRFSLHLQTD	
seq2	VTNITLSTQH	YRIHRSDVEP	VKEKTTEKDI	FAKSITAVRN	SFISLSTSL	DRFSLHQQTD	
seq3	VTNITLSTQH	YRIHRSDVEP	VKEKTTEKDI	FAKSITAVRN	SFISLSTSL	DRFSLHQQTD	
seq4	VTKITLSPQN	FRIQKQET--	LKEKSTEKNS	LAKSILAVKN	HFIELRSKLS	ERFISHKNTE	
seq5	VTKITLSPQN	FRIQKQETTL	LKEKSTEKNS	LAKSILAVKN	HFIELRSKLS	ERFISHKNTE	
	IPTHFHGRGS	ASEGRAVLTS	KTVKDFMLQK	LNSLDIKGNA			
	IPTHFHGRGS	ASEGRAVLTS	KTVKDFMLQK	LNSLDIKGNA			
	IPTHFHGRGN	ASEGRAVLTS	KTVKDFMLQK	LNSLDIKGNA			
	SSATHFHGRGS	ASEGRAVLTN	KVVKDFMLQT	LNDIDIRGSA			
	SSATHFHGRGS	ASEGRAVLTN	KVVKDFMLQT	LNDIDIRGSA			

Parse Multiple Sequence Alignment Results

1. Slice part of the alignment; 2. change format

```
#!/usr/local/bin/perl
use strict;
use warnings;

use Bio::AlignIO;

my $in = Bio::AlignIO->new(-file => "myalignment.aln",
                           -format => "clustalw" );

my $out = Bio::AlignIO->new(-file => ">out.phylip" ,
                           -format => 'phylip');

while ( my $aln = $in->next_aln() ) {
    my $new_aln = $aln->slice(5,100);
    $out->write_aln($new_aln);
}
```

script5.pl

Methods Implemented in Bio::SimpleAlign

Modifier methods

add seq
remove seq
purge
sort alphabetically
sort by list
set new reference
uniq seq

Sequence selection methods

each seq
each alphabetically
each seq with id
get seq by pos
get seq by id
seq with features

Create new alignments

select
select noncont
slice
remove columns
remove gaps

Change sequences within the MSA

splice by seq pos
map chars
uppercase
cigar line
match line
gap line
all gap line
gap col matrix

match
unmatch

MSA attributes

id
accession
description
missing char
match char
gap char
symbol chars

Alignment descriptors

score

consensus string

consensus iupac
consensus meta

is flush

length

maxdisplayname length

max metaname length

num residues

num sequences

average percentage identity

percentage identity

overall percentage identity

Alignment positions

column from residue numbe

Sequence names

displayname

set displayname count

set displayname flat

set displayname normal

source

Exercise 1. Retrieve an E. coli genome from NCBI (Genbank accession NC_000913). Make a fasta file with 500bp upstream regions of all transcripts.
Hint: You can do this by modifying script1.pl of this lecture.

Exercise 2. Modify script4.pl, so that this script can take in a third parameter maximum eval, and only output HSP with eval below the cutoff.